

# Package ‘BNPdensity’

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**Type** Package

**Title** Ferguson-Klass Type Algorithm for Posterior Normalized Random Measures

**Version** 2023.3.8

**Description** Bayesian nonparametric density estimation modeling mixtures by a Ferguson-Klass type algorithm for posterior normalized random measures.

**License** GPL (>= 2)

**Encoding** UTF-8

**LazyData** yes

**ByteCompile** yes

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**Depends** R(>= 3.5.0)

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**NeedsCompilation** no

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## R topics documented:

BNPdensity-package . . . . .	3
acidity . . . . .	4
add . . . . .	5

as.mcmc.multNRMI . . . . .	5
asNumeric_no_warning . . . . .	6
comment_on_NRMI_type . . . . .	7
compute_optimal_clustering . . . . .	7
compute_thinning_grid . . . . .	8
convert_to_mcmc . . . . .	8
cpo.multNRMI . . . . .	9
cpo.NRMI1 . . . . .	9
cpo.NRMI2 . . . . .	10
dist_name_k_index_converter . . . . .	11
dt_ . . . . .	11
enzyme . . . . .	12
Enzyme1.out . . . . .	12
Enzyme2.out . . . . .	13
expected_number_of_components_Dirichlet . . . . .	13
expected_number_of_components_stable . . . . .	14
fill_sigmas . . . . .	15
galaxy . . . . .	15
Galaxy1.out . . . . .	16
Galaxy2.out . . . . .	16
give_kernel_name . . . . .	17
GOFplots . . . . .	17
GOFplots_censored . . . . .	18
GOFplots_noncensored . . . . .	19
grid_from_data . . . . .	19
grid_from_data_censored . . . . .	20
grid_from_data_noncensored . . . . .	20
is_censored . . . . .	21
is_semiparametric . . . . .	21
MixNRMI1 . . . . .	22
MixNRMI1cens . . . . .	26
MixNRMI2 . . . . .	30
MixNRMI2cens . . . . .	35
MixPY1 . . . . .	39
MixPY2 . . . . .	41
multMixNRMI1 . . . . .	43
multMixNRMI1cens . . . . .	45
multMixNRMI2 . . . . .	47
multMixNRMI2cens . . . . .	49
MvInv . . . . .	51
plot.multNRMI . . . . .	52
plot.NRMI1 . . . . .	53
plot.NRMI2 . . . . .	53
plot.PY1 . . . . .	54
plot.PY2 . . . . .	55
plotCDF_censored . . . . .	55
plotCDF_noncensored . . . . .	56
plotfit_censored . . . . .	56

plotfit_noncensored . . . . .	57
plotPDF_censored . . . . .	58
plotPDF_noncensored . . . . .	58
plot_clustering_and_CDF . . . . .	59
plot_prior_number_of_components . . . . .	59
pp_plot_censored . . . . .	60
pp_plot_noncensored . . . . .	61
print.multNRMI . . . . .	62
print.NRMI1 . . . . .	62
print.NRMI2 . . . . .	63
print.PY1 . . . . .	64
print.PY2 . . . . .	64
process_dist_name . . . . .	65
qq_plot_censored . . . . .	65
qq_plot_noncensored . . . . .	66
salinity . . . . .	67
summary.multNRMI . . . . .	67
summary.NRMI1 . . . . .	68
summary.NRMI2 . . . . .	69
summary.PY1 . . . . .	70
summary.PY2 . . . . .	70
summarytext . . . . .	71
traceplot . . . . .	72

<b>Index</b>	<b>73</b>
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BNPdensity-package	<i>Bayesian nonparametric density estimation</i>
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---

## Description

This package performs Bayesian nonparametric density estimation for exact and censored data via a normalized random measure mixture model. The package allows the user to specify the mixture kernel, the mixing normalized measure and the choice of performing fully nonparametric mixtures on locations and scales, or semiparametric mixtures on locations only with common scale parameter. Options for the kernels are: two kernels with support in the real line (gaussian and double exponential), two more kernels in the positive line (gamma and lognormal) and one with bounded support (beta). The options for the normalized random measures are members of the class of normalized generalized gamma, which include the Dirichlet process, the normalized inverse gaussian process and the normalized stable process. The type of censored data handled by the package is right, left and interval.

## Details

Package:	BNPdensity
Type:	Package
Version:	2016.10

Date: 2016-10-14  
License: GPL version 2 or later  
LazyLoad: yes

The package includes four main functions: `MixNRMI1`, `MixNRMI2`, `MixNRMI1cens` and `MixNRMI2cens` which implement semiparametric and fully nonparametric mixtures for exact data, and semiparametric and fully nonparametric mixtures for censored data respectively. Additionally, the package includes several other functions required for sampling from conditional distributions in the MCMC implementation. These functions are intended for internal use only.

### Author(s)

Barrios, E., Lijoi, A., Nieto-Barajas, L. E. and Prünster, I.; Contributor: Guillaume Kon Kam King.; Maintainer: Ernesto Barrios <ebarrios at itam.mx>

### References

Barrios, E., Lijoi, A., Nieto-Barajas, L. E. and Prünster, I. (2013). Modeling with Normalized Random Measure Mixture Models. *Statistical Science*. Vol. 28, No. 3, 313-334.

Kon Kam King, G., Arbel, J. and Prünster, I. (2016). Species Sensitivity Distribution revisited: a Bayesian nonparametric approach. In preparation.

### See Also

[MixNRMI1](#), [MixNRMI2](#), [MixNRMI1cens](#), [MixNRMI2cens](#)

### Examples

```
example(MixNRMI1)
example(MixNRMI2)
example(MixNRMI1cens)
example(MixNRMI2cens)
```

---

acidity

*Acidity Index Dataset*

---

### Description

Concerns an acidity index measured in a sample of 155 lakes in north-central Wisconsin.

### Format

A real vector with 155 observations.

**References**

Crawford, S. L., DeGroot, M. H., Kadane, J. B. and Small, M. J. (1992). Modeling lake chemistry distributions: approximate Bayesian methods for estimating a finite mixture model. *Technometrics*, 34, 441-453.

**Examples**

```
data(acidity)
hist(acidity)
```

---

add	<i>Add x and y</i>
-----	--------------------

---

**Description**

This is a helper function for use in Reduce() over a list of vectors

**Usage**

```
add(x, y)
```

**Arguments**

x	first argument of the sum
y	second argument of the sum

**Value**

x + y

---

as.mcmc.multNRMI	<i>Convert the output of multMixNRMI into a coda mcmc object</i>
------------------	------------------------------------------------------------------

---

**Description**

Convert the output of multMixNRMI into a coda mcmc object

**Usage**

```
## S3 method for class 'multNRMI'
as.mcmc(x, ..., thinning_to = 1000, ncores = parallel::detectCores())
```

**Arguments**

x	Output of multMixNRMI.
...	Further arguments to be passed to specific methods
thinning_to	Final length of the chain after thinning.
ncores	Specify the number of cores to use in the conversion

**Value**

a coda::mcmc object

**Examples**

```
data(acidity)
out <- multMixNRMI1(acidity, parallel = TRUE, Nit = 10, ncores = 2)
coda::as.mcmc(out, ncores = 2)
```

---

asNumeric\_no\_warning *If the function Rmpfr::asNumeric returns a warning about inefficiency, silence it.*

---

**Description**

The function Rmpfr::asNumeric prints the following warning: In asMethod(object) : coercing "mpfr1" via "mpfr" (inefficient). It is not clear how to avoid it nor how to silence it, hence this function. A cleaner solution may be available at: <https://stackoverflow.com/questions/4948361/how-do-i-save-warnings-and-errors-as-output-from-a-function/4952908#4952908>

**Usage**

```
asNumeric_no_warning(x)
```

**Arguments**

x	An object of class Rmpfr::mpfr1
---	---------------------------------

**Value**

a "numeric" number

---

comment\_on\_NRMI\_type *Comment on the NRMI process depending on the value of the parameters*

---

### Description

Comment on the NRMI process depending on the value of the parameters

### Usage

```
comment_on_NRMI_type(NRMI_param = list(Alpha = 1, Kappa = 0, Gamma = 0.4))
```

### Arguments

NRMI\_param      A named list of the form list("Alpha" = 1, "Kappa" = 0, "Gamma" = 0.4)

### Value

A string containing a comment on the NRMI process

### Examples

```
BNPdensity::comment_on_NRMI_type(list("Alpha" = 1, "Kappa" = 0, "Gamma" = 0.4))
BNPdensity::comment_on_NRMI_type(list("Alpha" = 1, "Kappa" = 0.1, "Gamma" = 0.4))
BNPdensity::comment_on_NRMI_type(list("Alpha" = 1, "Kappa" = 0.1, "Gamma" = 0.5))
```

---

compute\_optimal\_clustering  
*Compute the optimal clustering from an MCMC sample*

---

### Description

Summarizes the posterior on all possible clusterings by an optimal clustering where optimality is defined as minimizing the posterior expectation of a specific loss function, the Variation of Information or Binder's loss function. Computation can be lengthy for large datasets, because of the large size of the space of all clusterings.

### Usage

```
compute_optimal_clustering(fit, loss_type = "VI")
```

### Arguments

fit                The fitted object, obtained from one of the MixNRMIx functions

loss\_type        Defines the loss function to be used in the expected posterior loss minimization. Can be one of "VI" (Variation of Information), "B" (Binder's loss), "NVI" (Normalized Variation of Information) or "NID" (Normalized Information Distance). Defaults to "VI".

**Value**

A vector of integers with the same size as the data, indicating the allocation of each data point.

---

`compute_thinning_grid` *Compute the grid for thinning the MCMC chain*

---

**Description**

This function creates an real grid then rounds it. If the grid is fine enough, there is a risk that rounding ties, i.e. iteration which are kept twice. To avoid this, if the total number of iterations is smaller than twice the number of iterations desired after thinning, the chain is not thinned.

**Usage**

```
compute_thinning_grid(Nit, thinning_to = 10)
```

**Arguments**

<code>Nit</code>	Length of the MCMC chain
<code>thinning_to</code>	Desired number of iterations after thinning.

**Value**

an integer vector of the MCMC iterations retained.

---

`convert_to_mcmc` *Convert the output of multMixNRMI into a coda mcmc object*

---

**Description**

Convert the output of multMixNRMI into a coda mcmc object

**Usage**

```
convert_to_mcmc(fitlist, thinning_to = 1000, ncores = parallel::detectCores())
```

**Arguments**

<code>fitlist</code>	Output of multMixNRMI.
<code>thinning_to</code>	Final length of the chain after thinning.
<code>ncores</code>	Specify the number of cores to use in the conversion

**Value**

a coda::mcmc object



---

cpo.multNRMI	<i>Extract the Conditional Predictive Ordinates (CPOs) from a list of fitted objects</i>
--------------	------------------------------------------------------------------------------------------

---

**Description**

This function assumes that all chains have the same size. To allow for different chain sizes, care should be paid to proper weighting.

**Usage**

```
## S3 method for class 'multNRMI'
cpo(object, ...)
```

**Arguments**

object	A fit obtained through from the functions MixNRMI1/MixNRMI1cens
...	Further arguments to be passed to generic function, ignored at the moment

**Value**

A vector of Conditional Predictive Ordinates (CPOs)

**Examples**

```
data(acidity)
out <- multMixNRMI1(acidity, parallel = TRUE, Nit = 10, ncores = 2)
cpo(out)
```

---

cpo.NRMI1	<i>Extract the Conditional Predictive Ordinates (CPOs) from a fitted object</i>
-----------	---------------------------------------------------------------------------------

---

**Description**

Extract the Conditional Predictive Ordinates (CPOs) from a fitted object

**Usage**

```
## S3 method for class 'NRMI1'
cpo(object, ...)
```

**Arguments**

object	A fit obtained through from the functions MixNRMI1/MixNRMI1cens
...	Further arguments to be passed to generic function, ignored at the moment

**Value**

A vector of Conditional Predictive Ordinates (CPOs)

**Examples**

```
data(acidity)
out <- MixNRMI1(acidity, Nit = 50)
cpo(out)
```

---

cpo.NRMI2	<i>Extract the Conditional Predictive Ordinates (CPOs) from a fitted object</i>
-----------	---------------------------------------------------------------------------------

---

**Description**

Extract the Conditional Predictive Ordinates (CPOs) from a fitted object

**Usage**

```
## S3 method for class 'NRMI2'
cpo(object, ...)
```

**Arguments**

object	A fit obtained through from the function MixNRMI2/MixNRMI2cens
...	Further arguments to be passed to generic function, ignored at the moment

**Value**

A vector of Conditional Predictive Ordinates (CPOs)

**Examples**

```
data(acidity)
out <- MixNRMI2(acidity, Nit = 50)
cpo(out)
```

---

dist\_name\_k\_index\_converter  
*Convert distribution names to indices*

---

**Description**

Convert distribution names to indices

**Usage**

```
dist_name_k_index_converter(distname)
```

**Arguments**

distname            a character representing the distribution name. Allowed names are "normal", "gamma", "beta", "exponential", "double exponential", "lognormal", "half-Cauchy", "half-normal", "half-student", "uniform" and "truncated normal", or their common abbreviations "norm", "exp", "lnorm", "halfcauchy", "halfnorm", "half" and "unif".

**Value**

an index describing the distribution. 1 = Normal; 2 = Gamma; 3 = Beta; 4 = Double Exponential; 5 = Lognormal, 6 = Half-Cauchy, 7 = Half-normal, 8 = Half-Student, 9 = Uniform, 10 = Truncated normal

---

dt\_                            *Non-standard student-t density*

---

**Description**

Computes the density.

**Usage**

```
dt_(x, df, mean, sd)
```

**Arguments**

x                            Numeric vector. Data set to which the density is evaluated.  
df                            Numeric constant. Degrees of freedom (> 0, maybe non-integer)  
mean                        Numeric constant. Location parameter.  
sd                            Positive numeric constant. Scale parameter.  
## The function is currently defined as function(x, df, mean, sd) dt((x - mean) /  
sd, df, ncp = 0) / sd

**Details**

For internal use

---

enzyme

*Enzyme Dataset*

---

**Description**

Concerns the distribution of enzymatic activity in the blood, for an enzyme involved in the metabolism of carcinogenic substances, among a group of 245 unrelated individuals.

**Format**

A data frame with 244 observations on the following variable:

**list("enzyme")** A numeric vector.

**References**

Bechtel, Y. C., Bonaiti-Pellie, C., Poisson, N., Magnette, J. and Bechtel, P.R. (1993). A population and family study of N-acetyltransferase using caffeine urinary metabolites. Clin. Pharm. Therp., 54, 134-141.

**Examples**

```
data(enzyme)
hist(enzyme)
```

---

Enzyme1.out

*Fit of MixNRMII function to the enzyme dataset*

---

**Description**

This object contains the output when setting `set.seed(150520)` and running the function `Enzyme1.out <- MixNRMII(enzyme, Alpha = 1, Kappa = 0.007, Gama = 0.5, distr.k = "gamma", distr.p0 = "gamma", asigma = 1, bsigma = 1, Meps = 0.005, Nit = 5000, Pbi = 0.2)`

**Details**

See function `MixNRMII`

**Examples**

```
data(Enzyme1.out)
```

---

 Enzyme2.out

*Fit of MixNRM12 function to the enzyme dataset*


---

### Description

This object contains the output when setting `set.seed(150520)` and running the function `Enzyme2.out`

```
<- MixNRM12(enzyme, Alpha = 1, Kappa = 0.007, Gama = 0.5, distr.k = "gamma", distr.py0 = "gamma", distr.pz0 = "gamma", mu.pz0 = 1, sigma.pz0 = 1, Meps = 0.005, Nit = 5000, Pbi = 0.2)
```

See function `MixNRM12`

### Examples

```
data(Enzyme2.out)
```

---

 expected\_number\_of\_components\_Dirichlet

*Computes the expected number of components for a Dirichlet process.*


---

### Description

Computes the expected number of components for a Dirichlet process.

### Usage

```
expected_number_of_components_Dirichlet(
  n,
  Alpha,
  ntrunc = NULL,
  silence = TRUE
)
```

### Arguments

<code>n</code>	Number of data points
<code>Alpha</code>	Numeric constant. Total mass of the centering measure.
<code>ntrunc</code>	Level of truncation when computing the expectation. Defaults to <code>n</code> . If greater than <code>n</code> , it is fixed to <code>n</code> .
<code>silence</code>	Boolean. Whether to print the current calculation step for the Stable process, as the function can be long

**Value**

A real value which approximates the expected number of components

Reference: P. De Blasi, S. Favaro, A. Lijoi, R. H. Mena, I. Prünster, and M. Ruggiero, “Are Gibbs-type priors the most natural generalization of the Dirichlet process?,” IEEE Trans. Pattern Anal. Mach. Intell., vol. 37, no. 2, pp. 212–229, 2015.

**Examples**

```
expected_number_of_components_Dirichlet(100, 1.2)
```

---

```
expected_number_of_components_stable
```

*Computes the expected number of components for a stable process.*

---

**Description**

Computes the expected number of components for a stable process.

**Usage**

```
expected_number_of_components_stable(n, Gama, ntrunc = NULL)
```

**Arguments**

n	Number of data points
Gama	Numeric constant. $0 \leq \text{Gama} \leq 1$ .
ntrunc	Level of truncation when computing the expectation. Defaults to n. If greater than n, it is fixed to n.

**Value**

A real value of type mpfr1 which approximates the expected number of components

In spite of the high precision arithmetic packages used for in function, it can be numerically unstable for small values of Gama. This is because evaluating a sum with alternated signs, in the generalized factorial coefficients, is tricky. Reference: P. De Blasi, S. Favaro, A. Lijoi, R. H. Mena, I. Prünster, and M. Ruggiero, “Are gibbs-type priors the most natural generalization of the Dirichlet process?,” IEEE Trans. Pattern Anal. Mach. Intell., vol. 37, no. 2, pp. 212–229, 2015.

**Examples**

```
expected_number_of_components_stable(100, 0.8)
```

---

fill_sigmas	<i>Repeat the common scale parameter of a semiparametric model to match the dimension of the location parameters.</i>
-------------	-----------------------------------------------------------------------------------------------------------------------

---

**Description**

Repeat the common scale parameter of a semiparametric model to match the dimension of the location parameters.

**Usage**

```
fill_sigmas(semiparametric_fit)
```

**Arguments**

semiparametric\_fit  
The result of the fit, obtained through the function MixNRM1.

**Value**

an adequate list of vectors of sigmas

---

galaxy	<i>Galaxy Data Set</i>
--------	------------------------

---

**Description**

Velocities of 82 galaxies diverging from our own galaxy.

**Format**

A data frame with 82 observations on the following variable:

**list("velocity")** A numeric vector.

**References**

Roeder, K. (1990) "Density estimation with confidence sets exemplified by superclusters and voids in the galaxies". Journal of the American Statistical Association. 85, 617-624.

**Examples**

```
data(galaxy)
hist(galaxy)
```

---

Galaxy1.out

*Fit of MixNRMI1 function to the galaxy dataset*

---

### Description

This object contains the output when setting `set.seed(150520)` and running the function `MixNRMI1(galaxy, Alpha = 1, Kappa = 0.015, Gama = 0.5, distr.k = "normal", distr.p0 = "gamma", asigma = 1, bsigma = 1, delta = 7, Meps = 0.005, Nit = 5000, Pbi = 0.2)`

### Details

See function `MixNRMI1`.

### Examples

```
data(Galaxy1.out)
```

---

Galaxy2.out

*Fit of MixNRMI2 function to the galaxy dataset*

---

### Description

This object contains the output when setting `set.seed(150520)` and running the function `Enzyme2.out <- MixNRMI2(x, Alpha = 1, Kappa = 0.007, Gama = 0.5, distr.k = "gamma", distr.py0 = "gamma", distr.pz0 = "gamma", mu.pz0 = 1, sigma.pz0 = 1, Meps = 0.005, Nit = 5000, Pbi = 0.2)`

### Details

See function `MixNRMI2`.

### Examples

```
data(Galaxy2.out)
```



---

give_kernel_name	<i>Gives the kernel name from the integer code</i>
------------------	----------------------------------------------------

---

**Description**

This function is used in the print methods for MixNRMI1, MixNRMI2, MixNRMI1cens, MixNRMI2cens, and all the multMixNRMIx versions

**Usage**

```
give_kernel_name(distr.k)
```

**Arguments**

distr.k	The distribution name for the kernel. Allowed names are "normal", "gamma", "beta", "double exponential", "lognormal" or their common abbreviations "norm", "exp", or an integer number identifying the mixture kernel: 1 = Normal; 2 = Gamma; 3 = Beta; 4 = Double Exponential; 5 = Lognormal.
---------	------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------

**Value**

A character with the name of the distribution used as the kernel

**Examples**

```
BNPdensity:::give_kernel_name(4)
```

---

GOFplots	<i>Plot Goodness of fits graphical checks for censored data</i>
----------	-----------------------------------------------------------------

---

**Description**

Plot Goodness of fits graphical checks for censored data

**Usage**

```
GOFplots(fit, qq_plot = FALSE, thinning_to = 500)
```

**Arguments**

fit	The result of the fit, obtained through the function MixNRMI1 or MixNRMI2, MixMRMI1cens or MixMRMI2cens
qq_plot	Whether to compute the QQ-plot
thinning_to	How many iterations to compute the mean posterior quantiles

**Value**

A density plot, a cumulative density plot with the Turnbull cumulative distribution, a percentile-percentile plot, and potentially a quantile-quantile plot.

**Examples**

```
set.seed(150520)
data(salinity)
out <- MixNRMI1cens(salinity$left, salinity$right, extras = TRUE, Nit = 100)
GOFplots(out)
```

---

GOFplots\_censored      *Plot Goodness of fits graphical checks for censored data*

---

**Description**

Plot Goodness of fits graphical checks for censored data

**Usage**

```
GOFplots_censored(fit, qq_plot = FALSE, thinning_to = 500)
```

**Arguments**

fit	The result of the fit, obtained through the function MixNRMI1 or MixNRMI2, MixMRMI1cens or MixMRMI2cens
qq_plot	Whether to compute the QQ-plot
thinning_to	How many iterations to compute the mean posterior quantiles

**Value**

A density plot, a cumulative density plot with the Turnbull cumulative distribution, and a percentile-percentile plot.

**Examples**

```
set.seed(150520)
data(salinty)
out <- MixNRMI1cens(salinity$left, salinity$right, extras = TRUE, Nit = 100)
BNPdensity:::GOFplots_censored(out)
```

---

GOFplots\_noncensored *Plot Goodness of fits graphical checks for non censored data*

---

**Description**

Plot Goodness of fits graphical checks for non censored data

**Usage**

```
GOFplots_noncensored(fit, qq_plot = FALSE, thinning_to = 500)
```

**Arguments**

fit	The result of the fit, obtained through the function MixNRMI1 or MixNRMI2, MixMRMI1cens or MixMRMI2cens
qq_plot	Whether to compute the QQ-plot
thinning_to	How many iterations to compute the mean posterior quantiles

**Value**

A density plot with histogram, a cumulative density plot with the empirical cumulative distribution, and a percentile-percentile plot.

**Examples**

```
set.seed(150520)
data(acidity)
out <- MixNRMI1(acidity, extras = TRUE, Nit = 100)
BNPdensity:::GOFplots_noncensored(out)
```

---

grid\_from\_data *Create a plotting grid from censored or non-censored data.*

---

**Description**

Create a plotting grid from censored or non-censored data.

**Usage**

```
grid_from_data(data, npoints = 100)
```

**Arguments**

data	Input data from which to compute the grid.
npoints	Number of points on the grid.

**Value**

a vector containing the plotting grid

---

grid\_from\_data\_censored

*Create a plotting grid from censored data.*

---

**Description**

Create a plotting grid from censored data.

**Usage**

```
grid_from_data_censored(data, npoints = 100)
```

**Arguments**

data	Censored input data from which to compute the grid.
npoints	Number of points on the grid.

**Value**

a vector containing the plotting grid

---

grid\_from\_data\_noncensored

*Create a plotting grid from non-censored data.*

---

**Description**

Create a plotting grid from non-censored data.

**Usage**

```
grid_from_data_noncensored(data, npoints = 100)
```

**Arguments**

data	Non-censored input data from which to compute the grid.
npoints	Number of points on the grid.

**Value**

a vector containing the plotting grid

---

is_censored	<i>Test if the data is censored</i>
-------------	-------------------------------------

---

**Description**

Test if the data is censored

**Usage**

```
is_censored(dat)
```

**Arguments**

dat                   The dataset to be tested

**Value**

TRUE if the data is censored

**Examples**

```
data(salinity)
BNPdensity:::is_censored(salinity)
```

---

is_semiparametric	<i>Tests if a fit is a semi parametric or nonparametric model.</i>
-------------------	--------------------------------------------------------------------

---

**Description**

Tests if a fit is a semi parametric or nonparametric model.

**Usage**

```
is_semiparametric(fit)
```

**Arguments**

fit                   The result of the fit, obtained through the function MixNRMI1 or MixNRMI2.

**Value**

TRUE if the fit is a semiparametric model

**Examples**

```

set.seed(150520)
data(acidity)
x <- enzyme
out <- MixNRMI1(enzyme, extras = TRUE, Nit = 10)
BNPdensity:::is_semiparametric(out)

```

---

MixNRMI1

*Normalized Random Measures Mixture of Type I*


---

**Description**

Bayesian nonparametric estimation based on normalized measures driven mixtures for locations.

**Usage**

```

MixNRMI1(
  x,
  probs = c(0.025, 0.5, 0.975),
  Alpha = 1,
  Kappa = 0,
  Gama = 0.4,
  distr.k = "normal",
  distr.p0 = 1,
  asigma = 0.5,
  bsigma = 0.5,
  delta_S = 3,
  delta_U = 2,
  Meps = 0.01,
  Nx = 150,
  Nit = 1500,
  Pbi = 0.1,
  epsilon = NULL,
  printtime = TRUE,
  extras = TRUE,
  adaptive = FALSE
)

```

**Arguments**

x	Numeric vector. Data set to which the density is fitted.
probs	Numeric vector. Desired quantiles of the density estimates.
Alpha	Numeric constant. Total mass of the centering measure. See details.
Kappa	Numeric positive constant. See details.
Gama	Numeric constant. $0 \leq \text{Gama} \leq 1$ . See details.

distr.k	The distribution name for the kernel. Allowed names are "normal", "gamma", "beta", "double exponential", "lognormal" or their common abbreviations "norm", "exp", or an integer number identifying the mixture kernel: 1 = Normal; 2 = Gamma; 3 = Beta; 4 = Double Exponential; 5 = Lognormal.
distr.p0	The distribution name for the centering measure. Allowed names are "normal", "gamma", "beta", or their common abbreviations "norm", "exp", or an integer number identifying the centering measure: 1 = Normal; 2 = Gamma; 3 = Beta.
asigma	Numeric positive constant. Shape parameter of the gamma prior on the standard deviation of the mixture kernel distr.k.
bsigma	Numeric positive constant. Rate parameter of the gamma prior on the standard deviation of the mixture kernel distr.k.
delta_S	Numeric positive constant. Metropolis-Hastings proposal variation coefficient for sampling sigma.
delta_U	Numeric positive constant. Metropolis-Hastings proposal variation coefficient for sampling the latent U.
Meps	Numeric constant. Relative error of the jump sizes in the continuous component of the process. Smaller values imply larger number of jumps.
Nx	Integer constant. Number of grid points for the evaluation of the density estimate.
Nit	Integer constant. Number of MCMC iterations.
Pbi	Numeric constant. Burn-in period proportion of Nit.
epsilon	Numeric constant. Extension to the evaluation grid range. See details.
printtime	Logical. If TRUE, prints out the execution time.
extras	Logical. If TRUE, gives additional objects: means, weights and Js.
adaptive	Logical. If TRUE, uses an adaptive MCMC strategy to sample the latent U (adaptive delta_U).

## Details

This generic function fits a normalized random measure (NRMI) mixture model for density estimation (James et al. 2009). Specifically, the model assumes a normalized generalized gamma (NGG) prior for the locations (means) of the mixture kernel and a parametric prior for the common smoothing parameter sigma, leading to a semiparametric mixture model.

The details of the model are:

$$\begin{aligned}
 X_i | Y_i, \sigma &\sim k(\cdot | Y_i, \sigma) \\
 Y_i | P &\sim P, \quad i = 1, \dots, n \\
 P &\sim \text{NGG}(\text{Alpha}, \text{Kappa}, \text{Gama}; P_\emptyset) \\
 \sigma &\sim \text{Gamma}(\text{asigma}, \text{bsigma})
 \end{aligned}$$

where  $X_i$ 's are the observed data,  $Y_i$ 's are latent (location) variables, sigma is the smoothing parameter, k is a parametric kernel parameterized in terms of mean and standard deviation, (Alpha, Kappa, Gama;  $P_\emptyset$ ) are the parameters of the NGG prior with  $P_\emptyset$  being the centering measure whose parameters are assigned vague hyper prior distributions, and (asigma,bsigma) are the hyper-parameters of the gamma prior on the smoothing parameter sigma. In particular: NGG(Alpha,

$1, \theta; P_{\theta}$ ) defines a Dirichlet process;  $NGG(1, \text{Kappa}, 1/2; P_{\theta})$  defines a Normalized inverse Gaussian process; and  $NGG(1, \theta, \text{Gama}; P_{\theta})$  defines a normalized stable process.

The evaluation grid ranges from  $\min(x) - \text{epsilon}$  to  $\max(x) + \text{epsilon}$ . By default  $\text{epsilon} = \text{sd}(x)/4$ .

### Value

The function returns a MixNRMI1 object. It is based on a list with the following components:

xx	Numeric vector. Evaluation grid.
qx	Numeric array. Matrix of dimension $N_x \times (\text{length}(\text{probs}) + 1)$ with the posterior mean and the desired quantiles input in probs.
cpo	Numeric vector of length(x) with conditional predictive ordinates.
R	Numeric vector of length(Nit*(1-Pbi)) with the number of mixtures components (clusters).
S	Numeric vector of length(Nit*(1-Pbi)) with the values of common standard deviation sigma.
U	Numeric vector of length(Nit*(1-Pbi)) with the values of the latent variable U.
Allocs	List of length(Nit*(1-Pbi)) with the clustering allocations.
means	List of length(Nit*(1-Pbi)) with the cluster means (locations). Only if extras = TRUE.
weights	List of length(Nit*(1-Pbi)) with the mixture weights. Only if extras = TRUE.
Js	List of length(Nit*(1-Pbi)) with the unnormalized weights (jump sizes). Only if extras = TRUE.
Nm	Integer constant. Number of jumps of the continuous component of the unnormalized process.
Nx	Integer constant. Number of grid points for the evaluation of the density estimate.
Nit	Integer constant. Number of MCMC iterations.
Pbi	Numeric constant. Burn-in period proportion of Nit.
procTime	Numeric vector with execution time provided by proc.time function.
distr.k	Integer corresponding to the kernel chosen for the mixture
data	Data used for the fit
NRMI_params	A named list with the parameters of the NRMI process

### Warning

The function is computing intensive. Be patient.

### Author(s)

Barrios, E., Kon Kam King, G., Lijoi, A., Nieto-Barajas, L.E. and Pruenster, I.



## References

- 1.- Barrios, E., Lijoi, A., Nieto-Barajas, L. E. and Prünster, I. (2013). Modeling with Normalized Random Measure Mixture Models. *Statistical Science*. Vol. 28, No. 3, 313-334.
- 2.- James, L.F., Lijoi, A. and Prünster, I. (2009). Posterior analysis for normalized random measure with independent increments. *Scand. J. Statist* 36, 76-97.

## See Also

[MixNRMI2](#), [MixNRMI1cens](#), [MixNRMI2cens](#), [multMixNRMI1](#)

## Examples

```
### Example 1
## Not run:
# Data
data(acidity)
x <- acidity
# Fitting the model under default specifications
out <- MixNRMI1(x)
# Plotting density estimate + 95% credible interval
plot(out)
### Example 2
set.seed(150520)
data(enzyme)
x <- enzyme
Enzyme1.out <- MixNRMI1(x, Alpha = 1, Kappa = 0.007, Gama = 0.5,
  distr.k = "gamma", distr.p0 = "gamma",
  asigma = 1, bsigma = 1, Meps=0.005,
  Nit = 5000, Pbi = 0.2)

attach(Enzyme1.out)
# Plotting density estimate + 95% credible interval
plot(Enzyme1.out)
# Plotting number of clusters
par(mfrow = c(2, 1))
plot(R, type = "l", main = "Trace of R")
hist(R, breaks = min(R - 0.5):max(R + 0.5), probability = TRUE)
# Plotting sigma
par(mfrow = c(2, 1))
plot(S, type = "l", main = "Trace of sigma")
hist(S, nclass = 20, probability = TRUE, main = "Histogram of sigma")
# Plotting u
par(mfrow = c(2, 1))
plot(U, type = "l", main = "Trace of U")
hist(U, nclass = 20, probability = TRUE, main = "Histogram of U")
# Plotting cpo
par(mfrow = c(2, 1))
plot(cpo, main = "Scatter plot of CPO's")
boxplot(cpo, horizontal = TRUE, main = "Boxplot of CPO's")
print(paste("Average log(CPO)=", round(mean(log(cpo)), 4)))
print(paste("Median log(CPO)=", round(median(log(cpo)), 4)))
```

```

detach()

## End(Not run)

### Example 3
## Do not run
# set.seed(150520)
# data(galaxy)
# x <- galaxy
# Galaxy1.out <- MixNRMI1(x, Alpha = 1, Kappa = 0.015, Gama = 0.5,
#                         distr.k = "normal", distr.p0 = "gamma",
#                         asigma = 1, bsigma = 1, delta = 7, Meps=0.005,
#                         Nit = 5000, Pbi = 0.2)

# The output of this run is already loaded in the package
# To show results run the following
# Data
data(galaxy)
x <- galaxy
data(Galaxy1.out)
attach(Galaxy1.out)
# Plotting density estimate + 95% credible interval
plot(Galaxy1.out)
# Plotting number of clusters
par(mfrow = c(2, 1))
plot(R, type = "l", main = "Trace of R")
hist(R, breaks = min(R - 0.5):max(R + 0.5), probability = TRUE)
# Plotting sigma
par(mfrow = c(2, 1))
plot(S, type = "l", main = "Trace of sigma")
hist(S, nclass = 20, probability = TRUE, main = "Histogram of sigma")
# Plotting u
par(mfrow = c(2, 1))
plot(U, type = "l", main = "Trace of U")
hist(U, nclass = 20, probability = TRUE, main = "Histogram of U")
# Plotting cpo
par(mfrow = c(2, 1))
plot(cpo, main = "Scatter plot of CPO's")
boxplot(cpo, horizontal = TRUE, main = "Boxplot of CPO's")
print(paste("Average log(CPO)=", round(mean(log(cpo)), 4)))
print(paste("Median log(CPO)=", round(median(log(cpo)), 4)))
detach()

```

## Description

Bayesian nonparametric estimation based on normalized measures driven mixtures for locations.

**Usage**

```

MixNRMIIcens(
  xleft,
  xright,
  probs = c(0.025, 0.5, 0.975),
  Alpha = 1,
  Kappa = 0,
  Gama = 0.4,
  distr.k = "normal",
  distr.p0 = "normal",
  asigma = 0.5,
  bsigma = 0.5,
  delta_S = 3,
  delta_U = 2,
  Meps = 0.01,
  Nx = 150,
  Nit = 1500,
  Pbi = 0.1,
  epsilon = NULL,
  printtime = TRUE,
  extras = TRUE,
  adaptive = FALSE
)

```

**Arguments**

<code>xleft</code>	Numeric vector. Lower limit of interval censoring. For exact data the same as <code>xright</code>
<code>xright</code>	Numeric vector. Upper limit of interval censoring. For exact data the same as <code>xleft</code> .
<code>probs</code>	Numeric vector. Desired quantiles of the density estimates.
<code>Alpha</code>	Numeric constant. Total mass of the centering measure. See details.
<code>Kappa</code>	Numeric positive constant. See details.
<code>Gama</code>	Numeric constant. $0 \leq \text{Gama} \leq 1$ . See details.
<code>distr.k</code>	The distribution name for the kernel. Allowed names are "normal", "gamma", "beta", "double exponential", "lognormal" or their common abbreviations "norm", "exp", or an integer number identifying the mixture kernel: 1 = Normal; 2 = Gamma; 3 = Beta; 4 = Double Exponential; 5 = Lognormal.
<code>distr.p0</code>	The distribution name for the centering measure. Allowed names are "normal", "gamma", "beta", or their common abbreviations "norm", "exp", or an integer number identifying the centering measure: 1 = Normal; 2 = Gamma; 3 = Beta.
<code>asigma</code>	Numeric positive constant. Shape parameter of the gamma prior on the standard deviation of the mixture kernel <code>distr.k</code> .
<code>bsigma</code>	Numeric positive constant. Rate parameter of the gamma prior on the standard deviation of the mixture kernel <code>distr.k</code> .

delta_S	Numeric positive constant. Metropolis-Hastings proposal variation coefficient for sampling sigma.
delta_U	Numeric positive constant. Metropolis-Hastings proposal variation coefficient for sampling the latent U.
Meps	Numeric constant. Relative error of the jump sizes in the continuous component of the process. Smaller values imply larger number of jumps.
Nx	Integer constant. Number of grid points for the evaluation of the density estimate.
Nit	Integer constant. Number of MCMC iterations.
Pbi	Numeric constant. Burn-in period proportion of Nit.
epsilon	Numeric constant. Extension to the evaluation grid range. See details.
printtime	Logical. If TRUE, prints out the execution time.
extras	Logical. If TRUE, gives additional objects: means, weights and Js.
adaptive	Logical. If TRUE, uses an adaptive MCMC strategy to sample the latent U (adaptive delta_U).

## Details

This generic function fits a normalized random measure (NRM) mixture model for density estimation (James et al. 2009) with censored data. Specifically, the model assumes a normalized generalized gamma (NGG) prior for the locations (means) of the mixture kernel and a parametric prior for the common smoothing parameter sigma, leading to a semiparametric mixture model.

This function coincides with `MixNRMI` when the lower (xleft) and upper (xright) censoring limits correspond to the same exact value.

The details of the model are:

$$X_i|Y_i, \sigma \sim k(\cdot|Y_i, \sigma)$$

$$Y_i|P \sim P, \quad i = 1, \dots, n$$

$$P \sim \text{NGG}(\text{Alpha}, \text{Kappa}, \text{Gama}; P\_0)$$

$$\sigma \sim \text{Gamma}(\text{asigma}, \text{bsigma})$$

where  $X_i$ 's are the observed data,  $Y_i$ 's are latent (location) variables, sigma is the smoothing parameter, k is a parametric kernel parameterized in terms of mean and standard deviation, (Alpha, Kappa, Gama; P\_0) are the parameters of the NGG prior with P\_0 being the centering measure whose parameters are assigned vague hyper prior distributions, and (asigma,bsigma) are the hyper-parameters of the gamma prior on the smoothing parameter sigma. In particular: `NGG(Alpha, 1, 0; P_0)` defines a Dirichlet process; `NGG(1, Kappa, 1/2; P_0)` defines a Normalized inverse Gaussian process; and `NGG(1, 0, Gama; P_0)` defines a normalized stable process.

The evaluation grid ranges from  $\min(x) - \text{epsilon}$  to  $\max(x) + \text{epsilon}$ . By default  $\text{epsilon} = \text{sd}(x)/4$ .

**Value**

The function returns a list with the following components:

xx	Numeric vector. Evaluation grid.
qx	Numeric array. Matrix of dimension $N_x \times (\text{length}(\text{probs}) + 1)$ with the posterior mean and the desired quantiles input in probs.
cpo	Numeric vector of length(x) with conditional predictive ordinates.
R	Numeric vector of length( $N_{it}*(1-P_{bi})$ ) with the number of mixtures components (clusters).
S	Numeric vector of length( $N_{it}*(1-P_{bi})$ ) with the values of common standard deviation sigma.
U	Numeric vector of length( $N_{it}*(1-P_{bi})$ ) with the values of the latent variable U.
Allocs	List of length( $N_{it}*(1-P_{bi})$ ) with the clustering allocations.
means	List of length( $N_{it}*(1-P_{bi})$ ) with the cluster means (locations). Only if extras = TRUE.
weights	List of length( $N_{it}*(1-P_{bi})$ ) with the mixture weights. Only if extras = TRUE.
Js	List of length( $N_{it}*(1-P_{bi})$ ) with the unnormalized weights (jump sizes). Only if extras = TRUE.
Nm	Integer constant. Number of jumps of the continuous component of the unnormalized process.
Nx	Integer constant. Number of grid points for the evaluation of the density estimate.
Nit	Integer constant. Number of MCMC iterations.
Pbi	Numeric constant. Burn-in period proportion of Nit.
procTime	Numeric vector with execution time provided by proc.time function.
distr.k	Integer corresponding to the kernel chosen for the mixture
data	Data used for the fit
NRMI_params	A named list with the parameters of the NRMI process

**Warning**

The function is computing intensive. Be patient.

**Author(s)**

Barrios, E., Kon Kam King, G. and Nieto-Barajas, L.E.

**References**

- 1.- Barrios, E., Lijoi, A., Nieto-Barajas, L. E. and Prünster, I. (2013). Modeling with Normalized Random Measure Mixture Models. *Statistical Science*. Vol. 28, No. 3, 313-334.
- 2.- James, L.F., Lijoi, A. and Prünster, I. (2009). Posterior analysis for normalized random measure with independent increments. *Scand. J. Statist* 36, 76-97.
- 3.- Kon Kam King, G., Arbel, J. and Prünster, I. (2016). Species Sensitivity Distribution revisited: a Bayesian nonparametric approach. In preparation.

**See Also**

[MixNRMI2](#), [MixNRMI1cens](#), [MixNRMI2cens](#), [multMixNRMI1](#)

**Examples**

```
### Example 1
## Not run:
# Data
data(acidity)
x <- acidity
# Fitting the model under default specifications
out <- MixNRMI1cens(x, x)
# Plotting density estimate + 95% credible interval
plot(out)

## End(Not run)

## Not run:
### Example 2
# Data
data(salinity)
# Fitting the model under default specifications
out <- MixNRMI1cens(xleft = salinity$left, xright = salinity$right, Nit = 5000)
# Plotting density estimate + 95% credible interval
attach(out)
plot(out)
# Plotting number of clusters
par(mfrow = c(2, 1))
plot(R, type = "l", main = "Trace of R")
hist(R, breaks = min(R - 0.5):max(R + 0.5), probability = TRUE)
detach()

## End(Not run)
```

---

MixNRMI2

*Normalized Random Measures Mixture of Type II*

---

**Description**

Bayesian nonparametric estimation based on normalized measures driven mixtures for locations and scales.

**Usage**

```
MixNRMI2(
  x,
  probs = c(0.025, 0.5, 0.975),
```

```

Alpha = 1,
Kappa = 0,
Gama = 0.4,
distr.k = "normal",
distr.py0 = "normal",
distr.pz0 = "gamma",
mu.pz0 = 3,
sigma.pz0 = sqrt(10),
delta_S = 4,
kappa = 2,
delta_U = 2,
Meps = 0.01,
Nx = 150,
Nit = 1500,
Pbi = 0.1,
epsilon = NULL,
printtime = TRUE,
extras = TRUE,
adaptive = FALSE
)

```

### Arguments

x	Numeric vector. Data set to which the density is fitted.
probs	Numeric vector. Desired quantiles of the density estimates.
Alpha	Numeric constant. Total mass of the centering measure. See details.
Kappa	Numeric positive constant. See details.
Gama	Numeric constant. $0 \leq Gama \leq 1$ . See details.
distr.k	The distribution name for the kernel. Allowed names are "normal", "gamma", "beta", "double exponential", "lognormal" or their common abbreviations "norm", "exp", or an integer number identifying the mixture kernel: 1 = Normal; 2 = Gamma; 3 = Beta; 4 = Double Exponential; 5 = Lognormal.
distr.py0	The distribution name for the centering measure for locations. Allowed names are "normal", "gamma", "beta", or their common abbreviations "norm", "exp", or an integer number identifying the centering measure for locations: 1 = Normal; 2 = Gamma; 3 = Beta.
distr.pz0	The distribution name for the centering measure for scales. Allowed names are "gamma", or an integer number identifying the centering measure for scales: 2 = Gamma. For more options use <a href="#">MixNRM12cens</a> .
mu.pz0	Numeric constant. Prior mean of the centering measure for scales.
sigma.pz0	Numeric constant. Prior standard deviation of the centering measure for scales.
delta_S	Numeric positive constant. Metropolis-Hastings proposal variation coefficient for sampling the scales.
kappa	Numeric positive constant. Metropolis-Hastings proposal variation coefficient for sampling the location parameters.

delta_U	Numeric positive constant. Metropolis-Hastings proposal variation coefficient for sampling the latent U. If 'adaptive=TRUE', 'delta_U' is the starting value for the adaptation.
Meps	Numeric constant. Relative error of the jump sizes in the continuous component of the process. Smaller values imply larger number of jumps.
Nx	Integer constant. Number of grid points for the evaluation of the density estimate.
Nit	Integer constant. Number of MCMC iterations.
Pbi	Numeric constant. Burn-in period proportion of Nit.
epsilon	Numeric constant. Extension to the evaluation grid range. See details.
printtime	Logical. If TRUE, prints out the execution time.
extras	Logical. If TRUE, gives additional objects: means, sigmas, weights and Js.
adaptive	Logical. If TRUE, uses an adaptive MCMC strategy to sample the latent U (adaptive delta_U).

### Details

This generic function fits a normalized random measure (NRM) mixture model for density estimation (James et al. 2009). Specifically, the model assumes a normalized generalized gamma (NGG) prior for both, locations (means) and standard deviations, of the mixture kernel, leading to a fully nonparametric mixture model.

The details of the model are:

$$\begin{aligned}
 X_i | Y_i, Z_i &\sim k(\cdot | Y_i, Z_i) \\
 (Y_i, Z_i) | P &\sim P, i = 1, \dots, n \\
 P &\sim \text{NGG}(\text{Alpha}, \text{Kappa}, \text{Gama}; P_\theta)
 \end{aligned}$$

where,  $X_i$ 's are the observed data,  $(Y_i, Z_i)$ 's are bivariate latent (location and scale) vectors,  $k$  is a parametric kernel parameterized in terms of mean and standard deviation,  $(\text{Alpha}, \text{Kappa}, \text{Gama}; P_\theta)$  are the parameters of the NGG prior with a bivariate  $P_\theta$  being the centering measure with independent components, that is,  $P_\theta(Y, Z) = P_\theta(Y) * P_\theta(Z)$ . The parameters of  $P_\theta(Y)$  are assigned vague hyper prior distributions and  $(\mu.pz\theta, \sigma.pz\theta)$  are the hyper-parameters of  $P_\theta(Z)$ . In particular,  $\text{NGG}(\text{Alpha}, 1, \theta; P_\theta)$  defines a Dirichlet process;  $\text{NGG}(1, \text{Kappa}, 1/2; P_\theta)$  defines a Normalized inverse Gaussian process; and  $\text{NGG}(1, \theta, \text{Gama}; P_\theta)$  defines a normalized stable process. The evaluation grid ranges from  $\min(x) - \text{epsilon}$  to  $\max(x) + \text{epsilon}$ . By default  $\text{epsilon} = \text{sd}(x)/4$ .

### Value

The function returns a list with the following components:

xx	Numeric vector. Evaluation grid.
qx	Numeric array. Matrix of dimension $Nx \times (\text{length}(\text{probs}) + 1)$ with the posterior mean and the desired quantiles input in probs.
cpo	Numeric vector of $\text{length}(x)$ with conditional predictive ordinates.
R	Numeric vector of $\text{length}(\text{Nit} * (1 - \text{Pbi}))$ with the number of mixtures components (clusters).



U	Numeric vector of length( $Nit*(1-Pbi)$ ) with the values of the latent variable U.
Allocs	List of length( $Nit*(1-Pbi)$ ) with the clustering allocations.
means	List of length( $Nit*(1-Pbi)$ ) with the cluster means (locations). Only if extras = TRUE.
sigmas	Numeric vector of length( $Nit*(1-Pbi)$ ) with the cluster standard deviations. Only if extras = TRUE.
weights	List of length( $Nit*(1-Pbi)$ ) with the mixture weights. Only if extras = TRUE.
Js	List of length( $Nit*(1-Pbi)$ ) with the unnormalized weights (jump sizes). Only if extras = TRUE.
Nm	Integer constant. Number of jumps of the continuous component of the unnormalized process.
delta_Us	List of length( $Nit*(1-Pbi)$ ) with the sequence of adapted delta_U used in the MH step for the latent variable U.
Nx	Integer constant. Number of grid points for the evaluation of the density estimate.
Nit	Integer constant. Number of MCMC iterations.
Pbi	Numeric constant. Burn-in period proportion of Nit.
procTime	Numeric vector with execution time provided by proc.time function.
distr.k	Integer corresponding to the kernel chosen for the mixture
data	Data used for the fit
NRMI_params	A named list with the parameters of the NRMI process

**Warning**

The function is computing intensive. Be patient.

**Author(s)**

Barrios, Kon Kam King, G., E., Lijoi, A., Nieto-Barajas, L.E. and Prünster, I.

**References**

- 1.- Barrios, E., Lijoi, A., Nieto-Barajas, L. E. and Prünster, I. (2013). Modeling with Normalized Random Measure Mixture Models. *Statistical Science*. Vol. 28, No. 3, 313-334.
- 2.- James, L.F., Lijoi, A. and Prünster, I. (2009). Posterior analysis for normalized random measure with independent increments. *Scand. J. Statist* 36, 76-97.
- 3.- Arbel, J., Kon Kam King, G., Lijoi, A., Nieto-Barajas, L.E. and Prünster, I. (2021). BNPdensity: a package for Bayesian Nonparametric density estimation using Normalised Random Measures with Independent Increments.. *Australian and New Zealand Journal of Statistics*, to appear

**See Also**

[MixNRMI2](#), [MixNRMI1cens](#), [MixNRMI2cens](#), [multMixNRMI1](#)

**Examples**

```

## Not run:
### Example 1
# Data
data(acidity)
x <- acidity
# Fitting the model under default specifications
out <- MixNRMI2(x)
# Plotting density estimate + 95% credible interval
plot(out)

## End(Not run)

### Example 2
## Do not run
# set.seed(150520)
# data(enzyme)
# x <- enzyme
# Enzyme2.out <- MixNRMI2(x, Alpha = 1, Kappa = 0.007, Gama = 0.5,
#                         distr.k = "gamma", distr.py0 = "gamma",
#                         distr.pz0 = "gamma", mu.pz0 = 1, sigma.pz0 = 1, Meps=0.005,
#                         Nit = 5000, Pbi = 0.2)
# The output of this run is already loaded in the package
# To show results run the following
# Data
data(enzyme)
x <- enzyme
data(Enzyme2.out)
attach(Enzyme2.out)
# Plotting density estimate + 95% credible interval
plot(Enzyme2.out)
# Plotting number of clusters
par(mfrow = c(2, 1))
plot(R, type = "l", main = "Trace of R")
hist(R, breaks = min(R - 0.5):max(R + 0.5), probability = TRUE)
# Plotting u
par(mfrow = c(2, 1))
plot(U, type = "l", main = "Trace of U")
hist(U, nclass = 20, probability = TRUE, main = "Histogram of U")
# Plotting cpo
par(mfrow = c(2, 1))
plot(cpo, main = "Scatter plot of CPO's")
boxplot(cpo, horizontal = TRUE, main = "Boxplot of CPO's")
print(paste("Average log(CPO)=", round(mean(log(cpo)), 4)))
print(paste("Median log(CPO)=", round(median(log(cpo)), 4)))
detach()

### Example 3
## Do not run
# set.seed(150520)
# data(galaxy)
# x <- galaxy

```

```

# Galaxy2.out <- MixNRMI2(x, Alpha = 1, Kappa = 0.015, Gama = 0.5,
#                         distr.k = "normal", distr.py0 = "gamma",
#                         distr.pz0 = "gamma", mu.pz0 = 1, sigma.pz0 = 1, Meps=0.005,
#                         Nit = 5000, Pbi = 0.2)
# The output of this run is already loaded in the package
# To show results run the following
# Data
data(galaxy)
x <- galaxy
data(Galaxy2.out)
attach(Galaxy2.out)
# Plotting density estimate + 95% credible interval
plot(Galaxy2.out)
# Plotting number of clusters
par(mfrow = c(2, 1))
plot(R, type = "l", main = "Trace of R")
hist(R, breaks = min(R - 0.5):max(R + 0.5), probability = TRUE)
# Plotting u
par(mfrow = c(2, 1))
plot(U, type = "l", main = "Trace of U")
hist(U, nclass = 20, probability = TRUE, main = "Histogram of U")
# Plotting cpo
par(mfrow = c(2, 1))
plot(cpo, main = "Scatter plot of CPO's")
boxplot(cpo, horizontal = TRUE, main = "Boxplot of CPO's")
print(paste("Average log(CPO)=", round(mean(log(cpo)), 4)))
print(paste("Median log(CPO)=", round(median(log(cpo)), 4)))
detach()

```

---

MixNRMI2cens

*Normalized Random Measures Mixture of Type II for censored data*


---

## Description

Bayesian nonparametric estimation based on normalized measures driven mixtures for locations and scales.

## Usage

```

MixNRMI2cens(
  xleft,
  xright,
  probs = c(0.025, 0.5, 0.975),
  Alpha = 1,
  Kappa = 0,
  Gama = 0.4,
  distr.k = "normal",
  distr.py0 = "normal",
  distr.pz0 = "gamma",

```

```

mu.pz0 = 3,
sigma.pz0 = sqrt(10),
delta_S = 4,
kappa = 2,
delta_U = 2,
Meps = 0.01,
Nx = 150,
Nit = 1500,
Pbi = 0.1,
epsilon = NULL,
printtime = TRUE,
extras = TRUE,
adaptive = FALSE
)

```

### Arguments

xleft	Numeric vector. Lower limit of interval censoring. For exact data the same as xright
xright	Numeric vector. Upper limit of interval censoring. For exact data the same as xleft.
probs	Numeric vector. Desired quantiles of the density estimates.
Alpha	Numeric constant. Total mass of the centering measure. See details.
Kappa	Numeric positive constant. See details.
Gama	Numeric constant. $0 \leq Gama \leq 1$ . See details.
distr.k	The distribution name for the kernel. Allowed names are "normal", "gamma", "beta", "double exponential", "lognormal" or their common abbreviations "norm", "exp", or an integer number identifying the mixture kernel: 1 = Normal; 2 = Gamma; 3 = Beta; 4 = Double Exponential; 5 = Lognormal.
distr.py0	The distribution name for the centering measure for locations. Allowed names are "normal", "gamma", "beta", or their common abbreviations "norm", "exp", or an integer number identifying the centering measure for locations: 1 = Normal; 2 = Gamma; 3 = Beta.
distr.pz0	The distribution name for the centering measure for scales. Allowed names are "gamma", "lognormal", "half-Cauchy", "half-normal", "half-student", "uniform" and "truncated normal", or their common abbreviations "norm", "exp", "lnorm", "halfcauchy", "halfnorm", "half" and "unif", or an integer number identifying the centering measure for scales: 2 = Gamma, 5 = Lognormal, 6 = Half Cauchy, 7 = Half Normal, 8 = Half Student-t, 9 = Uniform, 10 = Truncated Normal.
mu.pz0	Numeric constant. Prior mean of the centering measure for scales.
sigma.pz0	Numeric constant. Prior standard deviation of the centering measure for scales.
delta_S	Numeric positive constant. Metropolis-Hastings proposal variation coefficient for sampling the scales.
kappa	Numeric positive constant. Metropolis-Hastings proposal variation coefficient for sampling the location parameters.

delta_U	Numeric positive constant. Metropolis-Hastings proposal variation coefficient for sampling the latent U. If 'adaptive=TRUE', 'delta_U' is the starting value for the adaptation.
Meps	Numeric constant. Relative error of the jump sizes in the continuous component of the process. Smaller values imply larger number of jumps.
Nx	Integer constant. Number of grid points for the evaluation of the density estimate.
Nit	Integer constant. Number of MCMC iterations.
Pbi	Numeric constant. Burn-in period proportion of Nit.
epsilon	Numeric constant. Extension to the evaluation grid range. See details.
printtime	Logical. If TRUE, prints out the execution time.
extras	Logical. If TRUE, gives additional objects: means, sigmas, weights and Js.
adaptive	Logical. If TRUE, uses an adaptive MCMC strategy to sample the latent U (adaptive delta_U).

## Details

This generic function fits a normalized random measure (NRMI) mixture model for density estimation (James et al. 2009). Specifically, the model assumes a normalized generalized gamma (NGG) prior for both, locations (means) and standard deviations, of the mixture kernel, leading to a fully nonparametric mixture model.

The details of the model are:

$$\begin{aligned}
 X_i | Y_i, Z_i &\sim k(\cdot | Y_i, Z_i) \\
 (Y_i, Z_i) | P &\sim P, i = 1, \dots, n \\
 P &\sim \text{NGG}(\text{Alpha}, \text{Kappa}, \text{Gama}; P_\theta)
 \end{aligned}$$

where,  $X_i$ 's are the observed data,  $(Y_i, Z_i)$ 's are bivariate latent (location and scale) vectors,  $k$  is a parametric kernel parameterized in terms of mean and standard deviation,  $(\text{Alpha}, \text{Kappa}, \text{Gama}; P_\theta)$  are the parameters of the NGG prior with a bivariate  $P_\theta$  being the centering measure with independent components, that is,  $P_\theta(Y, Z) = P_\theta(Y) * P_\theta(Z)$ . The parameters of  $P_\theta(Y)$  are assigned vague hyper prior distributions and  $(\mu.pz\theta, \sigma.pz\theta)$  are the hyper-parameters of  $P_\theta(Z)$ . In particular,  $\text{NGG}(\text{Alpha}, 1, \theta; P_\theta)$  defines a Dirichlet process;  $\text{NGG}(1, \text{Kappa}, 1/2; P_\theta)$  defines a Normalized inverse Gaussian process; and  $\text{NGG}(1, \theta, \text{Gama}; P_\theta)$  defines a normalized stable process. The evaluation grid ranges from  $\min(x) - \text{epsilon}$  to  $\max(x) + \text{epsilon}$ . By default  $\text{epsilon} = \text{sd}(x)/4$ .

## Value

The function returns a list with the following components:

xx	Numeric vector. Evaluation grid.
qx	Numeric array. Matrix of dimension $Nx \times (\text{length}(\text{probs}) + 1)$ with the posterior mean and the desired quantiles input in probs.
cpo	Numeric vector of $\text{length}(x)$ with conditional predictive ordinates.
R	Numeric vector of $\text{length}(\text{Nit} * (1 - \text{Pbi}))$ with the number of mixtures components (clusters).

U	Numeric vector of length( $Nit*(1-Pbi)$ ) with the values of the latent variable U.
Allocs	List of length( $Nit*(1-Pbi)$ ) with the clustering allocations.
means	List of length( $Nit*(1-Pbi)$ ) with the cluster means (locations). Only if extras = TRUE.
sigmas	Numeric vector of length( $Nit*(1-Pbi)$ ) with the cluster standard deviations. Only if extras = TRUE.
weights	List of length( $Nit*(1-Pbi)$ ) with the mixture weights. Only if extras = TRUE.
Js	List of length( $Nit*(1-Pbi)$ ) with the unnormalized weights (jump sizes). Only if extras = TRUE.
Nm	Integer constant. Number of jumps of the continuous component of the unnormalized process.
delta_Us	List of length( $Nit*(1-Pbi)$ ) with the sequence of adapted delta_U used in the MH step for the latent variable U.
Nx	Integer constant. Number of grid points for the evaluation of the density estimate.
Nit	Integer constant. Number of MCMC iterations.
Pbi	Numeric constant. Burn-in period proportion of Nit.
procTime	Numeric vector with execution time provided by proc.time function.
distr.k	Integer corresponding to the kernel chosen for the mixture
data	Data used for the fit
NRMI_params	A named list with the parameters of the NRMI process

### Warning

The function is computing intensive. Be patient.

### Author(s)

Barrios, E., Kon Kam King, G. and Nieto-Barajas, L.E.

### References

- 1.- Barrios, E., Lijoi, A., Nieto-Barajas, L. E. and Prünster, I. (2013). Modeling with Normalized Random Measure Mixture Models. *Statistical Science*. Vol. 28, No. 3, 313-334.
- 2.- James, L.F., Lijoi, A. and Prünster, I. (2009). Posterior analysis for normalized random measure with independent increments. *Scand. J. Statist* 36, 76-97.
- 3.- Kon Kam King, G., Arbel, J. and Prünster, I. (2016). Species Sensitivity Distribution revisited: a Bayesian nonparametric approach. In preparation.

### See Also

[MixNRMI2](#), [MixNRMI1cens](#), [MixNRMI2cens](#), [multMixNRMI1](#)

**Examples**

```

## Not run:
### Example 1
# Data
data(acidity)
x <- acidity
# Fitting the model under default specifications
out <- MixNRMI2cens(x, x)
# Plotting density estimate + 95% credible interval
plot(out)

## End(Not run)

## Not run:
### Example 2
# Data
data(salinity)
# Fitting the model under special specifications
out <- MixNRMI2cens(
  xleft = salinity$left, xright = salinity$right, Nit = 5000, distr.pz0 = 10,
  mu.pz0 = 1, sigma.pz0 = 2
)
# Plotting density estimate + 95% credible interval
attach(out)
plot(out)
# Plotting number of clusters
par(mfrow = c(2, 1))
plot(R, type = "l", main = "Trace of R")
hist(R, breaks = min(R - 0.5):max(R + 0.5), probability = TRUE)
detach()

## End(Not run)

```

---

MixPY1

*Pitman-Yor process mixture of Type I*


---

**Description**

This function calls the PYdensity function from package BNPmix, to allow fitting a Pitman-Yor process mixture to the data.

**Usage**

```

MixPY1(
  x,
  probs = c(0.025, 0.5, 0.975),
  Alpha = 1,
  Gama = 0.4,

```

```

  asigma = 2,
  bsigma = 1/var(x),
  Nx = 100,
  Nit = 1500,
  Pbi = 0.5,
  epsilon = NULL,
  printtime = TRUE,
  extras = TRUE
)

```

### Arguments

x	Numeric vector. Data set to which the density is fitted.
probs	Numeric vector. Desired quantiles of the density estimates.
Alpha	Numeric constant. Total mass of the centering measure. See
Gama	Numeric constant. $0 \leq \text{Gama} \leq 1$ . See details.
asigma	Numeric positive constant. Shape parameter of the gamma prior on the standard deviation of the mixture kernel. Default value suggested by package BNPmix.
bsigma	Numeric positive constant. Rate parameter of the gamma prior on the standard deviation of the mixture kernel. Default value suggested by package BNPmix.
Nx	Integer constant. Number of grid points for the evaluation of the density estimate.
Nit	Integer constant. Number of MCMC iterations.
Pbi	Numeric constant. Burn-in period proportion of Nit.
epsilon	Numeric constant. Extension to the evaluation grid range. See details.
printtime	Logical. If TRUE, prints out the execution time.
extras	Logical. If TRUE, gives additional objects: means and weights

### Value

The function returns a MixPY1 object. It is based on a list with the following components:

xx	Numeric vector. Evaluation grid.
qx	Numeric array. Matrix of dimension $N_x \times (\text{length}(\text{probs}) + 1)$ with the posterior mean and the desired quantiles input in probs.
R	Numeric vector of length $(\text{Nit} * (1 - \text{Pbi}))$ with the number of mixtures components (clusters).
S	Numeric vector of length $(\text{Nit} * (1 - \text{Pbi}))$ with the values of common standard deviation sigma.
Allocs	List of length $(\text{Nit} * (1 - \text{Pbi}))$ with the clustering allocations.
means	List of length $(\text{Nit} * (1 - \text{Pbi}))$ with the cluster means (locations). Only if extras = TRUE.
weights	List of length $(\text{Nit} * (1 - \text{Pbi}))$ with the mixture weights. Only if extras = TRUE.
Nit	Integer constant. Number of MCMC iterations.



Pbi	Numeric constant. Burn-in period proportion of Nit.
distr.k	Integer corresponding to the kernel chosen for the mixture. Always 1, since the Pitman-Yor process is only written to work with Gaussian kernels.
data	Data used for the fit
PY_params	A named list with the parameters of the Pitman-Yor process

### Examples

```
# Data
data(acidity)
x <- acidity
# Fitting the model under default specifications
out <- MixPY1(x)
# Plotting density estimate + 95% credible interval
plot(out)
```

---

MixPY2

*Pitman-Yor process mixture of Type II*

---

### Description

This function calls the PYdensity function from package BNPmix, to allow fitting a Pitman-Yor process mixture to the data.

### Usage

```
MixPY2(
  x,
  probs = c(0.025, 0.5, 0.975),
  Alpha = 1,
  Gama = 0.4,
  asigma = 2,
  bsigma = 1/var(x),
  Nx = 100,
  Nit = 1500,
  Pbi = 0.5,
  epsilon = NULL,
  printtime = TRUE,
  extras = TRUE
)
```

### Arguments

x	Numeric vector. Data set to which the density is fitted.
probs	Numeric vector. Desired quantiles of the density estimates.
Alpha	Numeric constant. Total mass of the centering measure. See

Gama	Numeric constant. $0 \leq \text{Gama} \leq 1$ . See details.
asigma	Numeric positive constant. Shape parameter of the gamma prior on the standard deviation of the mixture kernel. Default value suggested by package BNPMix.
bsigma	Numeric positive constant. Rate parameter of the gamma prior on the standard deviation of the mixture kernel. Default value suggested by package BNPMix.
Nx	Integer constant. Number of grid points for the evaluation of the density estimate.
Nit	Integer constant. Number of MCMC iterations.
Pbi	Numeric constant. Burn-in period proportion of Nit.
epsilon	Numeric constant. Extension to the evaluation grid range. See details.
printtime	Logical. If TRUE, prints out the execution time.
extras	Logical. If TRUE, gives additional objects: means and weights

### Value

The function returns a MixPY2 object. It is based on a list with the following components:

xx	Numeric vector. Evaluation grid.
qx	Numeric array. Matrix of dimension $N_x \times (\text{length}(\text{probs}) + 1)$ with the posterior mean and the desired quantiles input in probs.
R	Numeric vector of length $(\text{Nit} * (1 - \text{Pbi}))$ with the number of mixtures components (clusters).
Allocs	List of length $(\text{Nit} * (1 - \text{Pbi}))$ with the clustering allocations.
means	List of length $(\text{Nit} * (1 - \text{Pbi}))$ with the cluster means (locations). Only if extras = TRUE.
sigmas	List of length $(\text{Nit} * (1 - \text{Pbi}))$ with the cluster standard deviations (scales). Only if extras = TRUE.
weights	List of length $(\text{Nit} * (1 - \text{Pbi}))$ with the mixture weights. Only if extras = TRUE.
Nit	Integer constant. Number of MCMC iterations.
Pbi	Numeric constant. Burn-in period proportion of Nit.
distr.k	Integer corresponding to the kernel chosen for the mixture. Always 1, since the Pitman-Yor process is only written to work with Gaussian kernels.
data	Data used for the fit
PY_params	A named list with the parameters of the Pitman-Yor process

### Examples

```
# Data
data(acidity)
x <- acidity
# Fitting the model under default specifications
out <- MixPY2(x)
# Plotting density estimate + 95% credible interval
plot(out)
```

---

multMixNRMI1

*Multiple chains of MixNRMI1*


---

## Description

Multiple chains of MixNRMI1

## Usage

```
multMixNRMI1(
  x,
  probs = c(0.025, 0.5, 0.975),
  Alpha = 1,
  Kappa = 0,
  Gama = 0.4,
  distr.k = "normal",
  distr.p0 = "normal",
  asigma = 0.5,
  bsigma = 0.5,
  delta_S = 3,
  delta_U = 2,
  Meps = 0.01,
  Nx = 150,
  Nit = 1500,
  Pbi = 0.1,
  epsilon = NULL,
  printtime = TRUE,
  extras = TRUE,
  adaptive = FALSE,
  nchains = 4,
  parallel = TRUE,
  ncores = parallel::detectCores()
)
```

## Arguments

x	Numeric vector. Data set to which the density is fitted.
probs	Numeric vector. Desired quantiles of the density estimates.
Alpha	Numeric constant. Total mass of the centering measure. See details.
Kappa	Numeric positive constant. See details.
Gama	Numeric constant. $0 \leq \text{Gama} \leq 1$ . See details.
distr.k	The distribution name for the kernel. Allowed names are "normal", "gamma", "beta", "double exponential", "lognormal" or their common abbreviations "norm", "exp", or an integer number identifying the mixture kernel: 1 = Normal; 2 = Gamma; 3 = Beta; 4 = Double Exponential; 5 = Lognormal.

distr.p0	The distribution name for the centering measure. Allowed names are "normal", "gamma", "beta", or their common abbreviations "norm", "exp", or an integer number identifying the centering measure: 1 = Normal; 2 = Gamma; 3 = Beta.
asigma	Numeric positive constant. Shape parameter of the gamma prior on the standard deviation of the mixture kernel <code>distr.k</code> .
bsigma	Numeric positive constant. Rate parameter of the gamma prior on the standard deviation of the mixture kernel <code>distr.k</code> .
delta_S	Numeric positive constant. Metropolis-Hastings proposal variation coefficient for sampling sigma.
delta_U	Numeric positive constant. Metropolis-Hastings proposal variation coefficient for sampling the latent U.
Meps	Numeric constant. Relative error of the jump sizes in the continuous component of the process. Smaller values imply larger number of jumps.
Nx	Integer constant. Number of grid points for the evaluation of the density estimate.
Nit	Integer constant. Number of MCMC iterations.
Pbi	Numeric constant. Burn-in period proportion of Nit.
epsilon	Numeric constant. Extension to the evaluation grid range. See details.
printtime	Logical. If TRUE, prints out the execution time.
extras	Logical. If TRUE, gives additional objects: means, weights and Js.
adaptive	Logical. If TRUE, uses an adaptive MCMC strategy to sample the latent U (adaptive delta_U).
nchains	The number of chains to run.
parallel	Whether to run the chains in parallel. Only works on UNIX-like systems as it rests on Fork parallelism
ncores	Number of cores for the parallel run. Defaults to <code>parallel::detectCores()</code> , i.e. the maximum number of cores detected by R on your system.

**Value**

a list containing the multiple fits.

**See Also**

[MixNRMI2](#), [MixNRMI1cens](#), [MixNRMI2cens](#)

**Examples**

```
data(acidity)
multMixNRMI1(acidity, parallel = TRUE, Nit = 10, ncores = 2)
```

---

multMixNRMI1cens      *Multiple chains of MixNRMI1cens*

---

## Description

Multiple chains of MixNRMI1cens

## Usage

```
multMixNRMI1cens(
  xleft,
  xright,
  probs = c(0.025, 0.5, 0.975),
  Alpha = 1,
  Kappa = 0,
  Gama = 0.4,
  distr.k = "normal",
  distr.p0 = "normal",
  asigma = 0.5,
  bsigma = 0.5,
  delta_S = 3,
  delta_U = 2,
  Meps = 0.01,
  Nx = 150,
  Nit = 1500,
  Pbi = 0.1,
  epsilon = NULL,
  printtime = TRUE,
  extras = TRUE,
  adaptive = FALSE,
  nchains = 4,
  parallel = TRUE,
  ncores = parallel::detectCores()
)
```

## Arguments

xleft	Numeric vector. Lower limit of interval censoring. For exact data the same as xright
xright	Numeric vector. Upper limit of interval censoring. For exact data the same as xleft.
probs	Numeric vector. Desired quantiles of the density estimates.
Alpha	Numeric constant. Total mass of the centering measure. See details.
Kappa	Numeric positive constant. See details.
Gama	Numeric constant. $0 \leq \text{Gama} \leq 1$ . See details.

<code>distr.k</code>	The distribution name for the kernel. Allowed names are "normal", "gamma", "beta", "double exponential", "lognormal" or their common abbreviations "norm", "exp", or an integer number identifying the mixture kernel: 1 = Normal; 2 = Gamma; 3 = Beta; 4 = Double Exponential; 5 = Lognormal.
<code>distr.p0</code>	The distribution name for the centering measure. Allowed names are "normal", "gamma", "beta", or their common abbreviations "norm", "exp", or an integer number identifying the centering measure: 1 = Normal; 2 = Gamma; 3 = Beta.
<code>asigma</code>	Numeric positive constant. Shape parameter of the gamma prior on the standard deviation of the mixture kernel <code>distr.k</code> .
<code>bsigma</code>	Numeric positive constant. Rate parameter of the gamma prior on the standard deviation of the mixture kernel <code>distr.k</code> .
<code>delta_S</code>	Numeric positive constant. Metropolis-Hastings proposal variation coefficient for sampling sigma.
<code>delta_U</code>	Numeric positive constant. Metropolis-Hastings proposal variation coefficient for sampling the latent U.
<code>Meps</code>	Numeric constant. Relative error of the jump sizes in the continuous component of the process. Smaller values imply larger number of jumps.
<code>Nx</code>	Integer constant. Number of grid points for the evaluation of the density estimate.
<code>Nit</code>	Integer constant. Number of MCMC iterations.
<code>Pbi</code>	Numeric constant. Burn-in period proportion of <code>Nit</code> .
<code>epsilon</code>	Numeric constant. Extension to the evaluation grid range. See details.
<code>printtime</code>	Logical. If TRUE, prints out the execution time.
<code>extras</code>	Logical. If TRUE, gives additional objects: means, weights and Js.
<code>adaptive</code>	Logical. If TRUE, uses an adaptive MCMC strategy to sample the latent U (adaptive <code>delta_U</code> ).
<code>nchains</code>	The number of chains to run.
<code>parallel</code>	Whether to run the chains in parallel. Only works on UNIX-like systems as it rests on Fork parallelism
<code>ncores</code>	Number of cores for the parallel run. Defaults to <code>parallel::detectCores()</code> , i.e. the maximum number of cores detected by R on your system.

**Value**

a list containing the multiple fits.

**See Also**

[MixNRMI2](#), [MixNRMI1cens](#), [MixNRMI2cens](#), [multMixNRMI1](#)

**Examples**

```
data(salinity)
multMixNRMI1cens(salinity$left, salinity$right, parallel = TRUE, Nit = 10, ncores = 2)
```

---

multMixNRMI2

*Multiple chains of MixNRMI2*


---

## Description

Multiple chains of MixNRMI2

## Usage

```
multMixNRMI2(
  x,
  probs = c(0.025, 0.5, 0.975),
  Alpha = 1,
  Kappa = 0,
  Gama = 0.4,
  distr.k = "normal",
  distr.py0 = "normal",
  distr.pz0 = "gamma",
  mu.pz0 = 3,
  sigma.pz0 = sqrt(10),
  delta_S = 4,
  kappa = 2,
  delta_U = 2,
  Meps = 0.01,
  Nx = 150,
  Nit = 1500,
  Pbi = 0.1,
  epsilon = NULL,
  printtime = TRUE,
  extras = TRUE,
  adaptive = FALSE,
  nchains = 4,
  parallel = FALSE,
  ncores = parallel::detectCores()
)
```

## Arguments

x	Numeric vector. Data set to which the density is fitted.
probs	Numeric vector. Desired quantiles of the density estimates.
Alpha	Numeric constant. Total mass of the centering measure. See details.
Kappa	Numeric positive constant. See details.
Gama	Numeric constant. $0 \leq Gama \leq 1$ . See details.
distr.k	The distribution name for the kernel. Allowed names are "normal", "gamma", "beta", "double exponential", "lognormal" or their common abbreviations "norm",

	"exp", or an integer number identifying the mixture kernel: 1 = Normal; 2 = Gamma; 3 = Beta; 4 = Double Exponential; 5 = Lognormal.
distr.py0	The distribution name for the centering measure for locations. Allowed names are "normal", "gamma", "beta", or their common abbreviations "norm", "exp", or an integer number identifying the centering measure for locations: 1 = Normal; 2 = Gamma; 3 = Beta.
distr.pz0	The distribution name for the centering measure for scales. Allowed names are "gamma", or an integer number identifying the centering measure for scales: 2 = Gamma. For more options use <a href="#">MixNRM12cens</a> .
mu.pz0	Numeric constant. Prior mean of the centering measure for scales.
sigma.pz0	Numeric constant. Prior standard deviation of the centering measure for scales.
delta_S	Numeric positive constant. Metropolis-Hastings proposal variation coefficient for sampling the scales.
kappa	Numeric positive constant. Metropolis-Hastings proposal variation coefficient for sampling the location parameters.
delta_U	Numeric positive constant. Metropolis-Hastings proposal variation coefficient for sampling the latent U. If 'adaptive=TRUE', 'delta_U' is the starting value for the adaptation.
Meps	Numeric constant. Relative error of the jump sizes in the continuous component of the process. Smaller values imply larger number of jumps.
Nx	Integer constant. Number of grid points for the evaluation of the density estimate.
Nit	Integer constant. Number of MCMC iterations.
Pbi	Numeric constant. Burn-in period proportion of Nit.
epsilon	Numeric constant. Extension to the evaluation grid range. See details.
printtime	Logical. If TRUE, prints out the execution time.
extras	Logical. If TRUE, gives additional objects: means, sigmas, weights and Js.
adaptive	Logical. If TRUE, uses an adaptive MCMC strategy to sample the latent U (adaptive delta_U).
nchains	The number of chains to run.
parallel	Whether to run the chains in parallel. Only works on UNIX-like systems as it rests on Fork parallelism
ncores	Number of cores for the parallel run. Defaults to parallel::detectCores(), i.e. the maximum number of cores detected by R on your system.

**Value**

a list containing the multiple fits.

**See Also**

[MixNRM12](#), [MixNRM1cens](#), [MixNRM12cens](#), [multMixNRM11](#)



## Examples

```
data(acidity)
multMixNRMI2(acidity, parallel = TRUE, Nit = 10, ncores = 2)
```

---

multMixNRMI2cens	<i>Multiple chains of MixNRMI2cens</i>
------------------	----------------------------------------

---

## Description

Multiple chains of MixNRMI2cens

## Usage

```
multMixNRMI2cens(  
  xleft,  
  xright,  
  probs = c(0.025, 0.5, 0.975),  
  Alpha = 1,  
  Kappa = 0,  
  Gama = 0.4,  
  distr.k = "normal",  
  distr.py0 = "normal",  
  distr.pz0 = "gamma",  
  mu.pz0 = 3,  
  sigma.pz0 = sqrt(10),  
  delta_S = 4,  
  kappa = 2,  
  delta_U = 2,  
  Meps = 0.01,  
  Nx = 150,  
  Nit = 1500,  
  Pbi = 0.1,  
  epsilon = NULL,  
  printtime = TRUE,  
  extras = TRUE,  
  adaptive = FALSE,  
  nchains = 4,  
  parallel = TRUE,  
  ncores = parallel::detectCores()  
)
```

## Arguments

xleft	Numeric vector. Lower limit of interval censoring. For exact data the same as xright
-------	--------------------------------------------------------------------------------------

xright	Numeric vector. Upper limit of interval censoring. For exact data the same as xleft.
probs	Numeric vector. Desired quantiles of the density estimates.
Alpha	Numeric constant. Total mass of the centering measure. See details.
Kappa	Numeric positive constant. See details.
Gama	Numeric constant. $0 \leq Gama \leq 1$ . See details.
distr.k	The distribution name for the kernel. Allowed names are "normal", "gamma", "beta", "double exponential", "lognormal" or their common abbreviations "norm", "exp", or an integer number identifying the mixture kernel: 1 = Normal; 2 = Gamma; 3 = Beta; 4 = Double Exponential; 5 = Lognormal.
distr.py0	The distribution name for the centering measure for locations. Allowed names are "normal", "gamma", "beta", or their common abbreviations "norm", "exp", or an integer number identifying the centering measure for locations: 1 = Normal; 2 = Gamma; 3 = Beta.
distr.pz0	The distribution name for the centering measure for scales. Allowed names are "gamma", "lognormal", "half-Cauchy", "half-normal", "half-student", "uniform" and "truncated normal", or their common abbreviations "norm", "exp", "lnorm", "halfcauchy", "halfnorm", "half" and "unif", or an integer number identifying the centering measure for scales: 2 = Gamma, 5 = Lognormal, 6 = Half Cauchy, 7 = Half Normal, 8 = Half Student-t, 9 = Uniform, 10 = Truncated Normal.
mu.pz0	Numeric constant. Prior mean of the centering measure for scales.
sigma.pz0	Numeric constant. Prior standard deviation of the centering measure for scales.
delta_S	Numeric positive constant. Metropolis-Hastings proposal variation coefficient for sampling the scales.
kappa	Numeric positive constant. Metropolis-Hastings proposal variation coefficient for sampling the location parameters.
delta_U	Numeric positive constant. Metropolis-Hastings proposal variation coefficient for sampling the latent U. If 'adaptive=TRUE', 'delta_U' is the starting value for the adaptation.
Meps	Numeric constant. Relative error of the jump sizes in the continuous component of the process. Smaller values imply larger number of jumps.
Nx	Integer constant. Number of grid points for the evaluation of the density estimate.
Nit	Integer constant. Number of MCMC iterations.
Pbi	Numeric constant. Burn-in period proportion of Nit.
epsilon	Numeric constant. Extension to the evaluation grid range. See details.
printtime	Logical. If TRUE, prints out the execution time.
extras	Logical. If TRUE, gives additional objects: means, sigmas, weights and Js.
adaptive	Logical. If TRUE, uses an adaptive MCMC strategy to sample the latent U (adaptive delta_U).
nchains	The number of chains to run.

parallel	Whether to run the chains in parallel. Only works on UNIX-like systems as it rests on Fork parallelism
ncores	Number of cores for the parallel run. Defaults to parallel::detectCores(), i.e. the maximum number of cores detected by R on your system.

**Value**

a list containing the multiple fits.

**See Also**

[MixNRMI2](#), [MixNRMI1cens](#), [MixNRMI2cens](#), [multMixNRMI1](#)

**Examples**

```
data(salinity)
## Not run:
multMixNRMI2cens(salinity$left, salinity$right, parallel = TRUE, Nit = 20, ncores = 2)
## End(Not run)
```

---

MvInv

*Invert jump heights function*


---

**Description**

Determines the jump heights of an increasing additive process by inverting the  $M(v)$  function. Use a truncation level based on expected moments of the NGG process ([thresholdGG](#)). For internal use.

**Usage**

```
MvInv(eps, u = 0.5, alpha = 1, kappa = 1, gama = 1/2, N = 3001)
```

**Arguments**

eps	Dummy argument kept for consistency with past versions of the functions
u	Real number. The value of the latent variable at the current step.
alpha	Numeric constant. Total mass of the centering measure.
kappa	Numeric positive constant.
gama	Numeric constant. Discount parameter of the NRMI process.

```

N          Number of steps in the discretization scheme for the grid inversion.

## The function has been optimised but it is morally defined as: function(eps, u
= 0.5, alpha = 1, kappa = 1, gama = 1 / 2, N = 3001) n <- length(w) v <- rep(NA,
n) x <- -log(seq(from = exp(-1e-05), to = exp(-10), length = N)) f <- alpha /
gamma(1 - gama) * x^(-(1 + gama)) * exp(-(u + kappa) * x) dx <- diff(x) h <-
(f[-1] + f[-N]) / 2 Mv <- rep(0, N) for (i in seq(N - 1, 1)) Mv[i] <- Mv[i + 1] +
dx[i] * h[i] for (j in seq(n)) v[j] <- x[which.min(Mv > w[j])] return(v)

```

---

```
plot.multNRMI
```

```
Plot the density estimate and the 95% credible interval
```

---

### Description

The density estimate is the mean posterior density computed on the data points.

### Usage

```
## S3 method for class 'multNRMI'
plot(x, ...)
```

### Arguments

```
x          An object of class multNRMI
...        Further arguments to be passed to generic functions, ignored at the moment
```

### Value

A graph with the density estimate, the 95% credible interval. Includes a histogram if the data is non censored.

### Examples

```
data(salinity)
fit <- multMixNRMI2cens(salinity$left, salinity$right, parallel = TRUE, Nit = 10, ncores = 2)
plot(fit)
```

---

plot.NRMI1	<i>Plot the density estimate and the 95% credible interval</i>
------------	----------------------------------------------------------------

---

**Description**

The density estimate is the mean posterior density computed on the data points.

**Usage**

```
## S3 method for class 'NRMI1'
plot(x, ...)
```

**Arguments**

x	A fitted object of class NRMI1
...	Further arguments to be passed to generic function, ignored at the moment

**Value**

A graph with the density estimate, the 95% credible interval and a histogram of the data

**Examples**

```
## Example for non censored data

data(acidity)
out <- MixNRMI1(acidity, Nit = 50)
plot(out)

## Example for censored data

data(salinity)
out <- MixNRMI1cens(salinity$left, salinity$right, Nit = 50)
plot(out)
```

---

plot.NRMI2	<i>Plot the density estimate and the 95% credible interval</i>
------------	----------------------------------------------------------------

---

**Description**

The density estimate is the mean posterior density computed on the data points.

**Usage**

```
## S3 method for class 'NRMI2'
plot(x, ...)
```

**Arguments**

x                    A fitted object of class NRMI2  
 ...                  Further arguments to be passed to generic function, ignored at the moment

**Value**

A graph with the density estimate, the 95% credible interval and a histogram of the data

**Examples**

```
## Example for non censored data

data(acidity)
out <- MixNRMI2(acidity, Nit = 20)
plot(out)

## Example for censored data

data(salinity)
out <- MixNRMI2cens(salinity$left, salinity$right, Nit = 20)
plot(out)
```

---

plot.PY1

---

*Plot the density estimate and the 95% credible interval*


---

**Description**

Plot the density estimate and the 95% credible interval

**Usage**

```
## S3 method for class 'PY1'
plot(x, ...)
```

**Arguments**

x                    A fitted object of class PY1  
 ...                  Further arguments to be passed to generic function, ignored at the moment

**Value**

A graph with the density estimate, the 95% credible interval and a histogram of the data

**Examples**

```
data(acidity)
out <- MixPY1(acidity, Nit = 50)
plot(out)
```

---

plot.PY2	<i>Plot the density estimate and the 95% credible interval</i>
----------	----------------------------------------------------------------

---

**Description**

Plot the density estimate and the 95% credible interval

**Usage**

```
## S3 method for class 'PY2'  
plot(x, ...)
```

**Arguments**

x	A fitted object of class PY2
...	Further arguments to be passed to generic function, ignored at the moment

**Value**

A graph with the density estimate, the 95% credible interval and a histogram of the data

**Examples**

```
data(acidity)  
out <- MixPY2(acidity, Nit = 50)  
plot(out)
```

---

plotCDF_censored	<i>Plot the Turnbull CDF and fitted CDF for censored data.</i>
------------------	----------------------------------------------------------------

---

**Description**

Plot the Turnbull CDF and fitted CDF for censored data.

**Usage**

```
plotCDF_censored(fit)
```

**Arguments**

fit	The result of the fit, obtained through the function MixNRMI1cens or MixNRMI2cens.
-----	------------------------------------------------------------------------------------

**Value**

Plot of the empirical and fitted CDF for non censored data.

## Examples

```
set.seed(150520)
data(salinity)
out <- MixNRM1cens(salinity$left, salinity$right, extras = TRUE, Nit = 100)
BNPdensity:::plotCDF_censored(out)
```

---

plotCDF\_noncensored     *Plot the empirical and fitted CDF for non censored data.*

---

## Description

Plot the empirical and fitted CDF for non censored data.

## Usage

```
plotCDF_noncensored(fit)
```

## Arguments

`fit`                    The result of the fit, obtained through the function `MixNRM1` or `MixNRM2`.

## Value

Plot of the empirical and fitted CDF for non censored data.

## Examples

```
set.seed(150520)
data(acidity)
out <- MixNRM1(acidity, extras = TRUE, Nit = 10)
BNPdensity:::plotCDF_noncensored(out)
```

---

plotfit\_censored         *Plot the density estimate and the 95% credible interval for censored data*

---

## Description

The density estimate is the mean posterior density computed on the data points. It is not possible to display a histogram for censored data.

## Usage

```
plotfit_censored(fit)
```



**Arguments**

fit                    A fitted object of class NRMI1cens or NRMI2cens

**Value**

A graph with the density estimate and the 95% credible interval

**Examples**

```
data(acidity)
out <- MixNRMI1(acidity, Nit = 50)
plot(out)
```

---

plotfit\_noncensored    *Plot the density estimate and the 95% credible interval for noncensored data*

---

**Description**

The density estimate is the mean posterior density computed on the data points.

**Usage**

```
plotfit_noncensored(fit)
```

**Arguments**

fit                    A fitted object of class NRMI1 or NRMI2

**Value**

A graph with the density estimate, the 95% credible interval and a histogram of the data

**Examples**

```
data(acidity)
out <- MixNRMI1(acidity, Nit = 50)
plot(out)
```

---

plotPDF\_censored      *Plot the density for censored data.*

---

**Description**

Plot the density for censored data.

**Usage**

```
plotPDF_censored(fit)
```

**Arguments**

`fit`                      The result of the fit, obtained through the function `MixNRMI1cens` or `MixNRMI2cens`.

**Value**

Plot of the density and a histogram for non censored data.

**Examples**

```
set.seed(150520)
data(salinity)
out <- MixNRMI1cens(xleft = salinity$left, xright = salinity$right, extras = TRUE, Nit = 100)
BNPdensity:::plotPDF_censored(out)
```

---

plotPDF\_noncensored      *Plot the density and a histogram for non censored data.*

---

**Description**

Plot the density and a histogram for non censored data.

**Usage**

```
plotPDF_noncensored(fit)
```

**Arguments**

`fit`                      The result of the fit, obtained through the function `MixNRMI1` or `MixNRMI2`.

**Value**

Plot of the density and a histogram for non censored data.

**Examples**

```
set.seed(150520)
data(acidity)
out <- MixNRM1(acidity, extras = TRUE, Nit = 100)
BNPdensity::plotPDF_noncensored(out)
```

---

plot\_clustering\_and\_CDF

*Plot the clustering and the Cumulative Distribution Function*

---

**Description**

This is a function to visualize the clustering induced by the BNP model. The data points are plotted with a color reflecting their cluster.

**Usage**

```
plot_clustering_and_CDF(fit, clustering, label_vector = NULL)
```

**Arguments**

fit	The fitted object, obtained from one of the MixNRM1x functions
clustering	A vector of integers with the same length as the data, representing the allocation variable for data each point.
label_vector	A vector of data labels to be plotted, to provide some identification to each point.

**Value**

A plot of the Cumulative Distribution Function (or Turnbull estimate for censored data) with data points whose color denotes the cluster allocation. For censored data, right or left censored data points are not represented, while interval censored data points are represented at the middle of the censoring interval.

---

plot\_prior\_number\_of\_components

*This plots the prior distribution on the number of components for the stable process. The Dirichlet process is provided for comparison.*

---

**Description**

This plots the prior distribution on the number of components for the stable process. The Dirichlet process is provided for comparison.

**Usage**

```
plot_prior_number_of_components(
  n,
  Gama,
  Alpha = 1,
  grid = NULL,
  silence = TRUE
)
```

**Arguments**

n	Number of data points
Gama	Numeric constant. $0 \leq \text{Gama} \leq 1$ .
Alpha	Numeric constant. Total mass of the centering measure for the Dirichlet process.
grid	Integer vector. Level of truncation when computing the expectation. Defaults to n. If greater than n, it is fixed to n.
silence	Boolean. Whether to print the current calculation step for the Stable process, as the function can be long

**Value**

A plot with the prior distribution on the number of components.

**Examples**

```
plot_prior_number_of_components(50, 0.4)
```

---

pp_plot_censored	<i>Plot the percentile-percentile graph for non censored data, using the Turnbull estimator the position of the percentiles.</i>
------------------	----------------------------------------------------------------------------------------------------------------------------------

---

**Description**

Plot the percentile-percentile graph for non censored data, using the Turnbull estimator the position of the percentiles.

**Usage**

```
pp_plot_censored(fit)
```

**Arguments**

fit	The result of the fit, obtained through the function MixNRMI1cens or MixNRMI2cens.
-----	------------------------------------------------------------------------------------

**Value**

Percentile-percentile graph using the Turnbull estimator

**Examples**

```
set.seed(150520)
data(salinity)
out <- MixNRM1cens(xleft = salinity$left, xright = salinity$right, extras = TRUE, Nit = 100)
BNPdensity::pp_plot_censored(out)
```

---

pp\_plot\_noncensored *Plot the percentile-percentile graph for non censored data.*

---

**Description**

Plot the percentile-percentile graph for non censored data.

**Usage**

```
pp_plot_noncensored(fit)
```

**Arguments**

fit                    The result of the fit, obtained through the function MixNRM1 or MixNRM2.

**Value**

Percentile-percentile plot for non censored data.

**Examples**

```
set.seed(150520)
data(acidity)
out <- MixNRM1(acidity, extras = TRUE, Nit = 100)
BNPdensity::pp_plot_noncensored(out)
```

---

```
print.multNRMI      S3 method for class 'multNRMI'
```

---

**Description**

S3 method for class 'multNRMI'

**Usage**

```
## S3 method for class 'multNRMI'
print(x, ...)
```

**Arguments**

```
x          An object of class multNRMI
...        Further arguments to be passed to generic functions, ignored at the moment
```

**Value**

A visualization of the important information about the object

**Examples**

```
data(salinity)
out <- multMixNRMI2cens(salinity$left, salinity$right, parallel = TRUE, Nit = 10, ncores = 2)
print(out)
```

---

```
print.NRMI1      S3 method for class 'MixNRMI1'
```

---

**Description**

S3 method for class 'MixNRMI1'

**Usage**

```
## S3 method for class 'NRMI1'
print(x, ...)
```

**Arguments**

```
x          A fitted object of class NRMI1
...        Further arguments to be passed to generic function, ignored at the moment
```

**Value**

A visualization of the important information about the object

**Examples**

```
## Example for non censored data

data(acidity)
out <- MixNRMI1(acidity, Nit = 50)
print(out)

## Example for censored data

data(salinity)
out <- MixNRMI1cens(salinity$left, salinity$right, Nit = 50)
print(out)
```

---

print.NRMI2	<i>S3 method for class 'MixNRMI2'</i>
-------------	---------------------------------------

---

**Description**

S3 method for class 'MixNRMI2'

**Usage**

```
## S3 method for class 'NRMI2'
print(x, ...)
```

**Arguments**

x	A fitted object of class NRMI2
...	Further arguments to be passed to generic function, ignored at the moment

**Value**

A visualization of the important information about the object

**Examples**

```
' ## Example for censored data
data(acidity)
out <- MixNRMI2(acidity, Nit = 20)
print(out)

data(salinity)
out <- MixNRMI2cens(salinity$left, salinity$right, Nit = 20)
print(out)
```

---

```
print.PY1          S3 method for class 'PY1'
```

---

**Description**

S3 method for class 'PY1'

**Usage**

```
## S3 method for class 'PY1'
print(x, ...)
```

**Arguments**

```
x          A fitted object of class PY1
...        Further arguments to be passed to generic function, ignored at the moment
```

**Value**

A visualization of the important information about the object

**Examples**

```
## Example for non censored data

data(acidity)
out <- MixPY1(acidity, Nit = 50)
print(out)
```

---

```
print.PY2          S3 method for class 'PY2'
```

---

**Description**

S3 method for class 'PY2'

**Usage**

```
## S3 method for class 'PY2'
print(x, ...)
```

**Arguments**

```
x          A fitted object of class PY2
...        Further arguments to be passed to generic function, ignored at the moment
```



**Value**

A visualization of the important information about the object

**Examples**

```
## Example for non censored data

data(acidity)
out <- MixPY2(acidity, Nit = 50)
print(out)
```

---

process_dist_name	<i>Process the distribution name argument into a distribution index</i>
-------------------	-------------------------------------------------------------------------

---

**Description**

This function is intended to help with compatibility with the previous versions of the package.

**Usage**

```
process_dist_name(distname)
```

**Arguments**

distname	Can be an integer or a distribution name. Allowed names are "normal", "gamma", "beta", "exponential", "lognormal", "half-Cauchy", "half-normal", "half-student", "uniform" and "truncated normal", or their common abbreviations "norm", "exp", "halfcauchy", "halfnorm", "half" and "unif".
----------	----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------

**Value**

an integer both if distname is an integer or a character

---

qq_plot_censored	<i>Plot the quantile-quantile graph for censored data.</i>
------------------	------------------------------------------------------------

---

**Description**

This function may be rather slow for many iterations/many data because it relies on numerical inversion of the mixture Cumulative Distribution Function. `set.seed(150520) data(salinity) out <- MixNRMI1cens(xleft = salinity$left, xright = salinity$right, extras = TRUE, Nit = 100) BNPdensity:::qq_plot_censored(out)`

**Usage**

```
qq_plot_censored(fit, thinning_to = 500)
```

**Arguments**

`fit`                The result of the fit, obtained through the function `MixNRMI1` or `MixNRMI2`, `MixMRMI1cens` or `MixMRMI2cens`

`thinning_to`        How many iterations to compute the mean posterior quantiles

**Value**

quantile-quantile plot for non censored data.

---

`qq_plot_noncensored`    *Plot the quantile-quantile graph for non censored data.*

---

**Description**

This function may be rather slow for many iterations/many data because it relies on numerical inversion of the mixture Cumulative Distribution Function.

**Usage**

```
qq_plot_noncensored(fit, thinning_to = 500)
```

**Arguments**

`fit`                The result of the fit, obtained through the function `MixNRMI1` or `MixNRMI2`, `MixMRMI1cens` or `MixMRMI2cens`

`thinning_to`        How many iterations to compute the mean posterior quantiles

**Value**

quantile-quantile plot for non censored data.

**Examples**

```
### Not run
# set.seed(150520)
# data(acidity)
# out <- MixNRMI1(acidity, extras = TRUE, Nit = 100)
# BNPdensity:::qq_plot_noncensored(out)
```

---

salinity

*Salinity tolerance*

---

### Description

72-hour acute salinity tolerance (LC50 values) of riverine macro-invertebrates.

### Format

A data frame with 108 observations on the following two variables:

**left** A numeric vector.

**right** A numeric vector.

### Source

fitdistrplus R-package

### References

Kefford, B.J., Nuggeoda, D., Metzeling, L., Fields, E. 2006. Validating species sensitivity distributions using salinity tolerance of riverine macroinvertebrates in the southern Murray-darling Basin (Victoria, Australia). Canadian Journal of Fisheries and Aquatic Science, 63, 1865-1877.

### Examples

```
data(salinity)
hist(salinity$left)
```

---

summary.multNRMI

*S3 method for class 'multNRMI'*

---

### Description

S3 method for class 'multNRMI'

### Usage

```
## S3 method for class 'multNRMI'
summary(object, number_of_clusters = FALSE, ...)
```

**Arguments**

object            A fitted object of class NRMI1cens  
 number\_of\_clusters  
                   Whether to compute the optimal number of clusters, which can be a time-consuming operation (see [compute\\_optimal\\_clustering](#))  
 ...                Further arguments to be passed to generic function, ignored at the moment

**Value**

Prints out the text for the summary S3 methods

**Examples**

```
data(salinity)
out <- multMixNRMI2cens(salinity$left, salinity$right, parallel = TRUE, Nit = 10, ncores = 2)
summary(out)
```

---

summary.NRMI1            *S3 method for class 'MixNRMI1'*

---

**Description**

S3 method for class 'MixNRMI1'

**Usage**

```
## S3 method for class 'NRMI1'
summary(object, number_of_clusters = FALSE, ...)
```

**Arguments**

object            A fitted object of class NRMI1  
 number\_of\_clusters  
                   Whether to compute the optimal number of clusters, which can be a time-consuming operation (see [compute\\_optimal\\_clustering](#))  
 ...                Further arguments to be passed to generic function, ignored at the moment

**Value**

Prints out the text for the summary S3 methods

## Examples

```
## Example for non censored data

data(acidity)
out <- MixNRMI1(acidity, Nit = 50)
summary(out)
```

---

summary.NRMI2

*S3 method for class 'MixNRMI2'*

---

## Description

S3 method for class 'MixNRMI2'

## Usage

```
## S3 method for class 'NRMI2'
summary(object, number_of_clusters = FALSE, ...)
```

## Arguments

object	A fitted object of class NRMI2
number_of_clusters	Whether to compute the optimal number of clusters, which can be a time-consuming operation (see <a href="#">compute_optimal_clustering</a> )
...	Further arguments to be passed to generic function, ignored at the moment

## Value

Prints out the text for the summary S3 methods

## Examples

```
data(acidity)
out <- MixNRMI2(acidity, Nit = 20)
summary(out)

data(salinity)
out <- MixNRMI2cens(salinity$left, salinity$right, Nit = 20)
summary(out)
```

summary.PY1

*S3 method for class 'PY1'*

---

**Description**

S3 method for class 'PY1'

**Usage**

```
## S3 method for class 'PY1'  
summary(object, number_of_clusters = FALSE, ...)
```

**Arguments**

object	A fitted object of class PY1
number_of_clusters	Whether to compute the optimal number of clusters, which can be a time-consuming operation (see <a href="#">compute_optimal_clustering</a> )
...	Further arguments to be passed to generic function, ignored at the moment

**Value**

Prints out the text for the summary S3 methods

**Examples**

```
## Example for non censored data  
  
data(acidity)  
out <- MixPY1(acidity, Nit = 50)  
summary(out)
```

---

summary.PY2*S3 method for class 'PY2'*

---

**Description**

S3 method for class 'PY2'

**Usage**

```
## S3 method for class 'PY2'  
summary(object, number_of_clusters = FALSE, ...)
```

**Arguments**

object            A fitted object of class PY2  
 number\_of\_clusters    Whether to compute the optimal number of clusters, which can be a time-consuming operation (see [compute\\_optimal\\_clustering](#))  
 ...                Further arguments to be passed to generic function, ignored at the moment

**Value**

Prints out the text for the summary S3 methods

**Examples**

```
## Example for non censored data

data(acidity)
out <- MixPY2(acidity, Nit = 50)
summary(out)
```

---

summarytext	<i>Common text for the summary S3 methods</i>
-------------	-----------------------------------------------

---

**Description**

Common text for the summary S3 methods

**Usage**

```
summarytext(
  fit,
  kernel_comment,
  BNP_process_comment,
  number_of_clusters = FALSE
)
```

**Arguments**

fit                NRMIX or PYx object  
 kernel\_comment    Text specific to the parametric and nonparametric nature of the model  
 BNP\_process\_comment    Text specific to the nonparametric process, NRMI or Pitman-Yor  
 number\_of\_clusters    Flag to decide whether to compute the optimal clustering

**Value**

Prints out the text for the summary S3 methods

---

traceplot	<i>Draw a traceplot for multiple chains</i>
-----------	---------------------------------------------

---

**Description**

This is a convenience function which works when coda is not yet loaded by the user. If coda is loaded, it gets masked. See also file multMixNRMI.R

**Usage**

```
traceplot(fitlist)
```

**Arguments**

fitlist            Output of multMixNRMI.

**Value**

A traceplot for multiple chains.



# Index

- \* **datasets**
    - acidity, [4](#)
    - enzyme, [12](#)
    - Enzyme1.out, [12](#)
    - Enzyme2.out, [13](#)
    - galaxy, [15](#)
    - Galaxy1.out, [16](#)
    - Galaxy2.out, [16](#)
    - salinity, [67](#)
  - \* **distribution**
    - MixNRMI1, [22](#)
    - MixNRMI1cens, [26](#)
    - MixNRMI2, [30](#)
    - MixNRMI2cens, [35](#)
  - \* **models**
    - MixNRMI1, [22](#)
    - MixNRMI1cens, [26](#)
    - MixNRMI2, [30](#)
    - MixNRMI2cens, [35](#)
  - \* **nonparametrics**
    - MixNRMI1, [22](#)
    - MixNRMI1cens, [26](#)
    - MixNRMI2, [30](#)
    - MixNRMI2cens, [35](#)
  - \* **package**
    - BNPdensity-package, [3](#)
- acidity, [4](#)  
add, [5](#)  
as.mcmc.multNRMI, [5](#)  
asNumeric\_no\_warning, [6](#)
- BNPdensity (BNPdensity-package), [3](#)  
BNPdensity-package, [3](#)
- comment\_on\_NRMI\_type, [7](#)  
compute\_optimal\_clustering, [7](#), [68–71](#)  
compute\_thinning\_grid, [8](#)  
convert\_to\_mcmc, [8](#)  
cpo.multNRMI, [9](#)
- cpo.NRMI1, [9](#)  
cpo.NRMI2, [10](#)
- dist\_name\_k\_index\_converter, [11](#)  
dt\_, [11](#)
- enzyme, [12](#)  
Enzyme1.out, [12](#)  
Enzyme2.out, [13](#)  
expected\_number\_of\_components\_Dirichlet, [13](#)  
expected\_number\_of\_components\_stable, [14](#)
- fill\_sigmas, [15](#)
- galaxy, [15](#)  
Galaxy1.out, [16](#)  
Galaxy2.out, [16](#)  
give\_kernel\_name, [17](#)  
GOFplots, [17](#)  
GOFplots\_censored, [18](#)  
GOFplots\_noncensored, [19](#)  
grid\_from\_data, [19](#)  
grid\_from\_data\_censored, [20](#)  
grid\_from\_data\_noncensored, [20](#)
- is\_censored, [21](#)  
is\_semiparametric, [21](#)
- MixNRMI1, [4](#), [22](#), [28](#)  
MixNRMI1cens, [4](#), [25](#), [26](#), [30](#), [33](#), [38](#), [44](#), [46](#), [48](#), [51](#)  
MixNRMI2, [4](#), [25](#), [30](#), [30](#), [33](#), [38](#), [44](#), [46](#), [48](#), [51](#)  
MixNRMI2cens, [4](#), [25](#), [30](#), [31](#), [33](#), [35](#), [38](#), [44](#), [46](#), [48](#), [51](#)
- MixPY1, [39](#)  
MixPY2, [41](#)  
multMixNRMI1, [25](#), [30](#), [33](#), [38](#), [43](#), [46](#), [48](#), [51](#)  
multMixNRMI1cens, [45](#)  
multMixNRMI2, [47](#)

multMixNRMI2cens, [49](#)  
MvInv, [51](#)

plot.multNRMI, [52](#)  
plot.NRMI1, [53](#)  
plot.NRMI2, [53](#)  
plot.PY1, [54](#)  
plot.PY2, [55](#)  
plot\_clustering\_and\_CDF, [59](#)  
plot\_prior\_number\_of\_components, [59](#)  
plotCDF\_censored, [55](#)  
plotCDF\_noncensored, [56](#)  
plotfit\_censored, [56](#)  
plotfit\_noncensored, [57](#)  
plotPDF\_censored, [58](#)  
plotPDF\_noncensored, [58](#)  
pp\_plot\_censored, [60](#)  
pp\_plot\_noncensored, [61](#)  
print.multNRMI, [62](#)  
print.NRMI1, [62](#)  
print.NRMI2, [63](#)  
print.PY1, [64](#)  
print.PY2, [64](#)  
process\_dist\_name, [65](#)

qq\_plot\_censored, [65](#)  
qq\_plot\_noncensored, [66](#)

salinity, [67](#)  
summary.multNRMI, [67](#)  
summary.NRMI1, [68](#)  
summary.NRMI2, [69](#)  
summary.PY1, [70](#)  
summary.PY2, [70](#)  
summarytext, [71](#)

thresholdGG, [51](#)  
traceplot, [72](#)